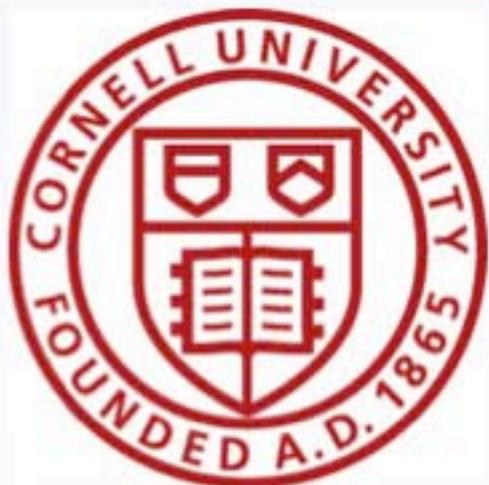


*International Workshop on Environmental Proteomics*  
Wednesday, January 20<sup>th</sup>, 2010

# **Targeted peptide quantification of *Dehalococcoides* proteins in a mixed culture during dehalorespiration**

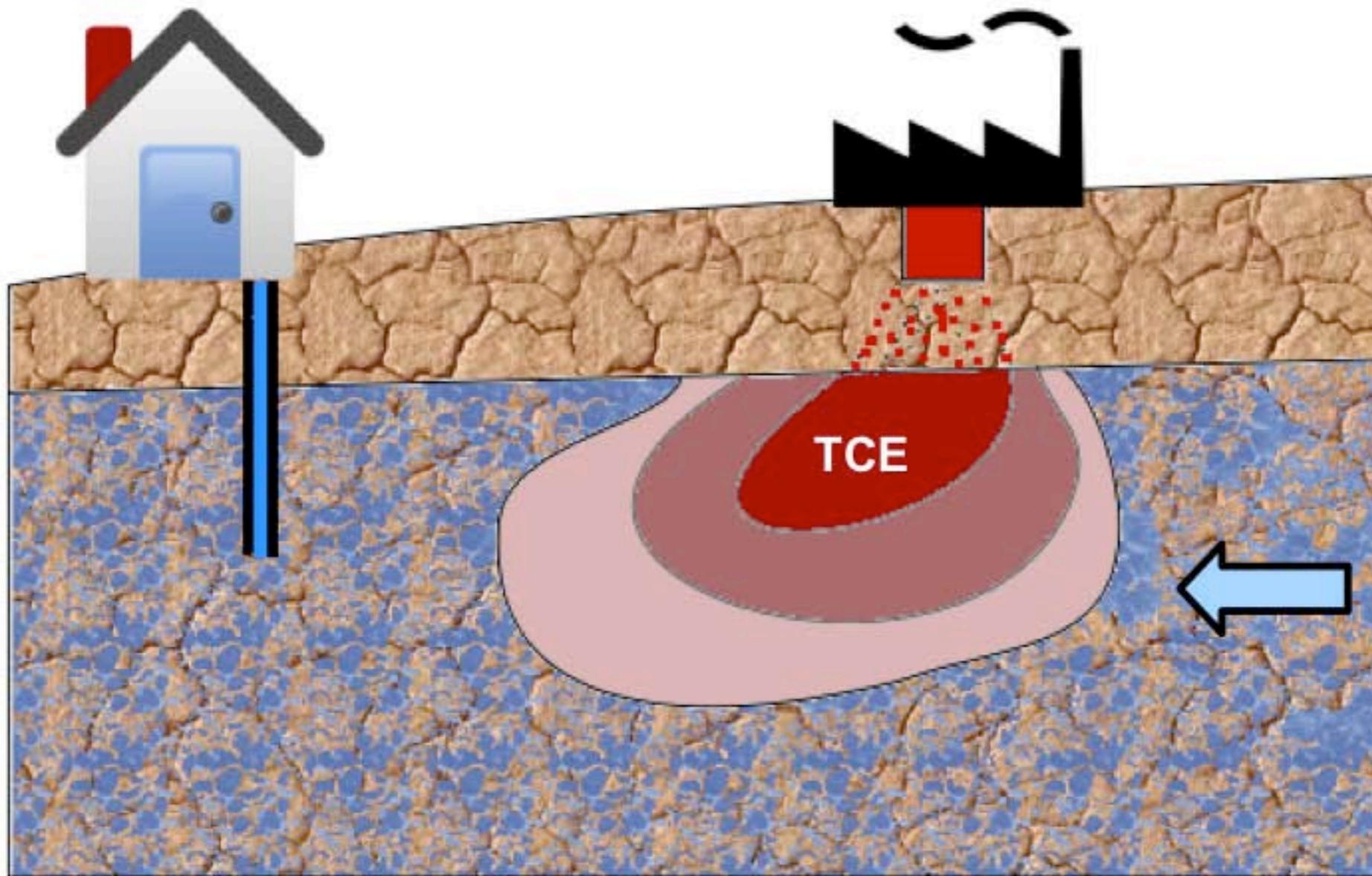
**Jeff Werner<sup>1</sup>, Celeste Ptak<sup>2</sup>,  
Sheng Zhang<sup>2</sup>, Ruth Richardson<sup>3</sup>**



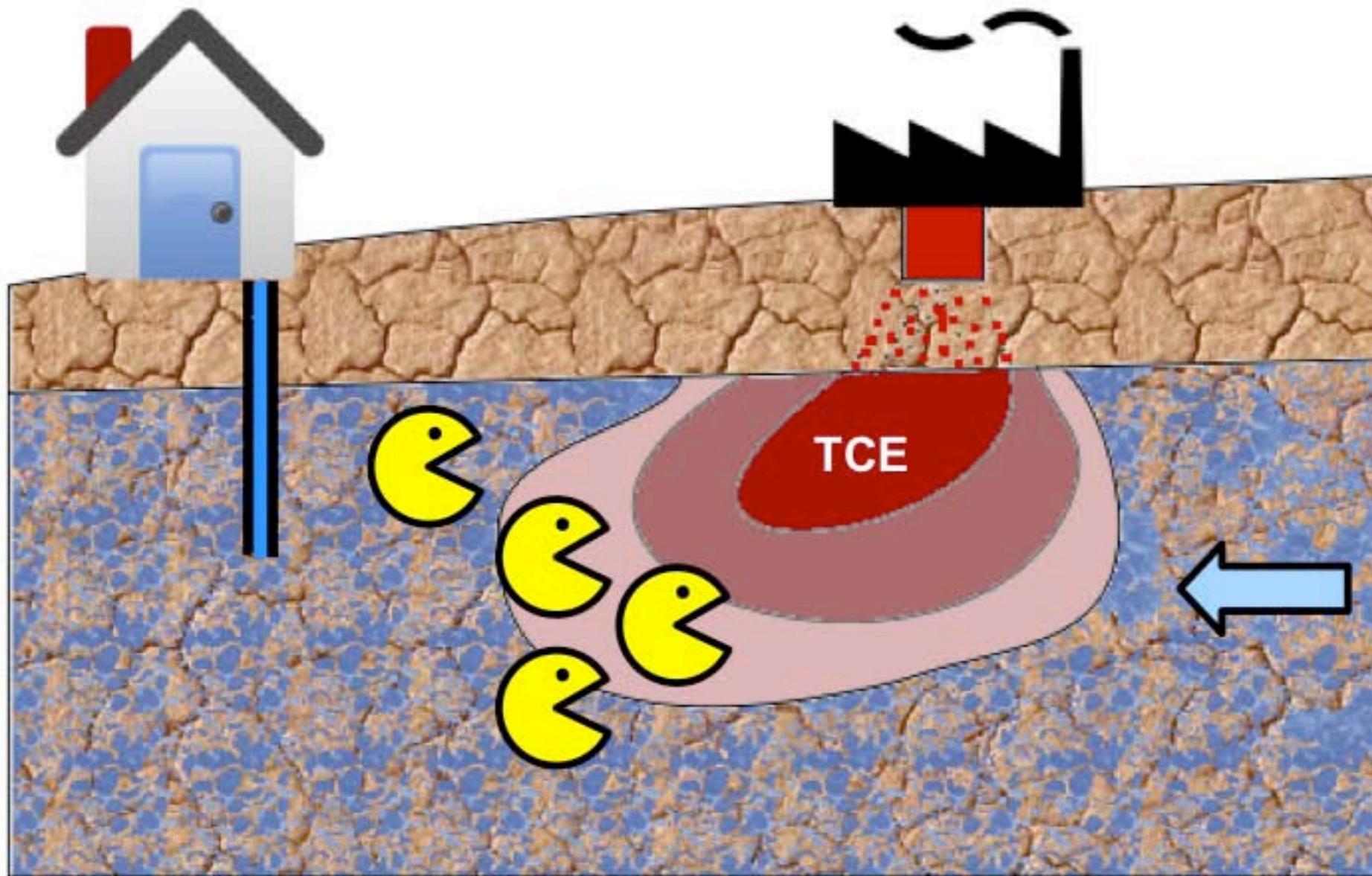
1. Dept. Biological and Environmental Engineering
2. Proteomics and Mass Spectrometry Facility
3. School of Civil and Environmental Engineering

Cornell University, Ithaca, NY

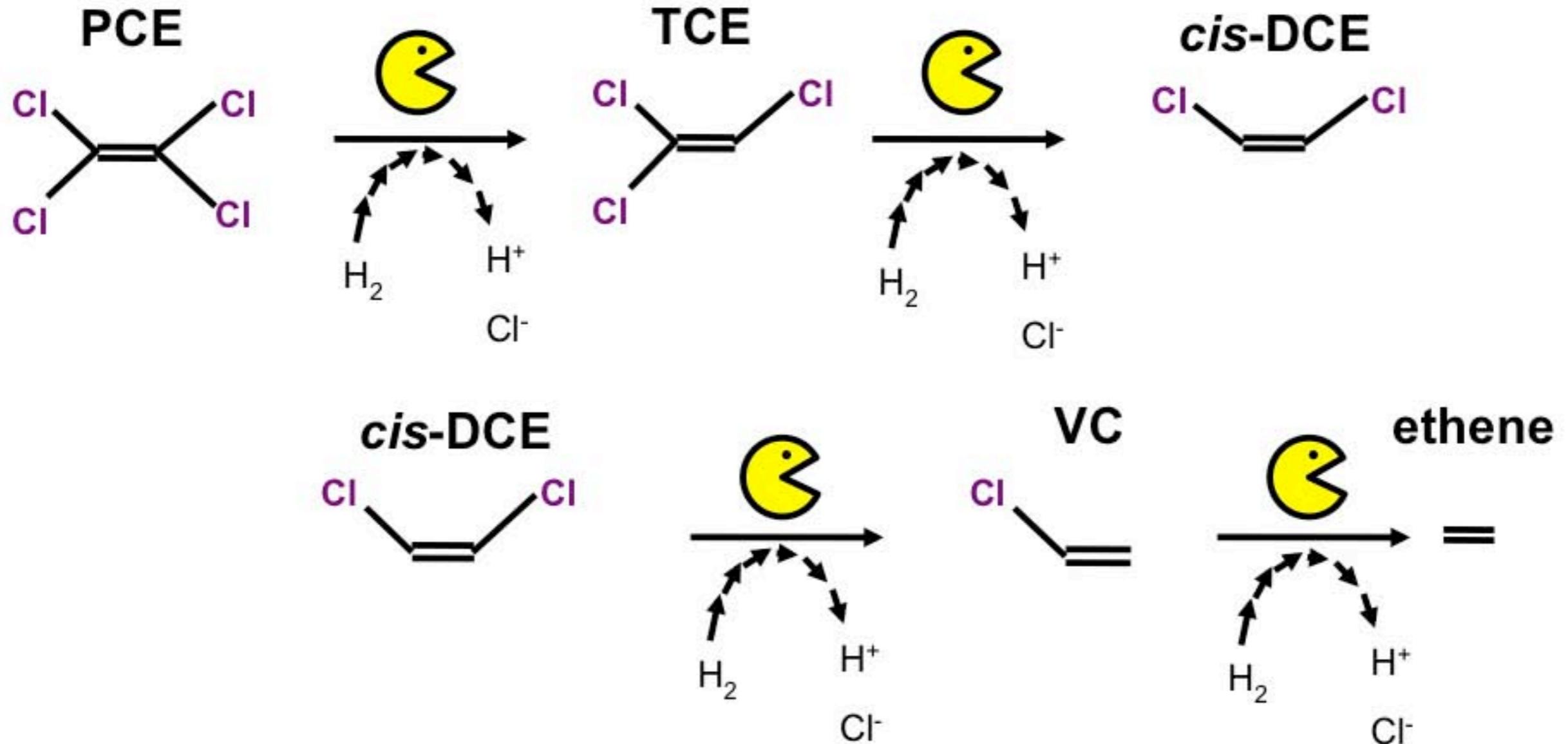
# Bioremediation



# Bioremediation



# Dehalorespiration



*Dehalococcoides* spp. are the only organisms known to do all these steps metabolically.

# Big-Picture Questions

- How can we assess dehalorespiration rates in the field?
- How can we tell if *Dehalococcoides* are healthy and active?

# **Mixed Cultures With Active *Dehalococcoides* spp.**

## **D2 (Richardson lab)**

- Complex mixed culture
- Maintained on PCE and Butyrate

## **KB1 (E Edwards lab, U Toronto)**

- Commercial mixed culture for field site bioaugmentation
- Enriched on TCE and Methanol

# **Proteins known to correlate with dehalorespiration activity**

- Reductive dehalogenases (eg TceA, PceA)
- Other oxidoreductases: (eg FdhA, HupA)

# Lots of shotgun data (more later)

## Sequence of TceA (from genome)

MSEKYHSTVTRRDFMKRLGLAGAGAGALGAAVLAENNLPH<sup>>95%</sup>E<sup>>50%</sup>FKDVDDL  
LSAGKALEGDHANKVNNHPWWVTTRDHEDPTCNIDWSLIK<sup>>95%</sup>RYSGWNN  
QGAYFLPEDYLSPTYTGR<sup>>95%</sup>R<sup>>50%</sup>HTIV<sup>>50%</sup>DSKLEIELQGK<sup>>95%</sup>KYR<sup>>50%</sup>DSAFIKSGIDWM  
KENIDPDYDPGELGYGDRREDALIYAATNGSHNCWENPLYGR<sup>>95%</sup>YEGSRP  
YLSM<sup>>95%</sup>RTMNGINGLHEFGHADIKTTNYPK<sup>>95%</sup>WEGTPEENLLIMRTAARYFGA  
SSVGAIKITDNVKKIFYAK<sup>>95%</sup>VQPFCLGPWYTITNMAEYIEYPVPVDNYAIPV  
FEDIPADQGHYSYK<sup>>95%</sup>RFGGDDKIAVPNALDNIFTYTIMLPEK<sup>>95%</sup>RFKYAHSIP  
MDPCSCIAAYPL<sup>>50%</sup>FTEVEAR<sup>>50%</sup>IQQFIAGLGYNSMGGGVEAWGPGSAFGNLS  
GLGEQSRV<sup>>50%</sup>SSIIEPRYGSNTK<sup>>50%</sup>GSLRMLTDLPLAPTKPIDAGIREFCKTCG  
ICAEHCPTQAISHEGPR<sup>>95%</sup>YDSPHWDCVSGYEGW<sup>>95%</sup>HLDYHKCINCTICEAVC  
PFFTMSNNSWVHNLVK<sup>>95%</sup>STVATTPVFNGFFKNMEGAFGYGPRY<sup>>95%</sup>SPSRDE  
WWASENPIRGASVDIF

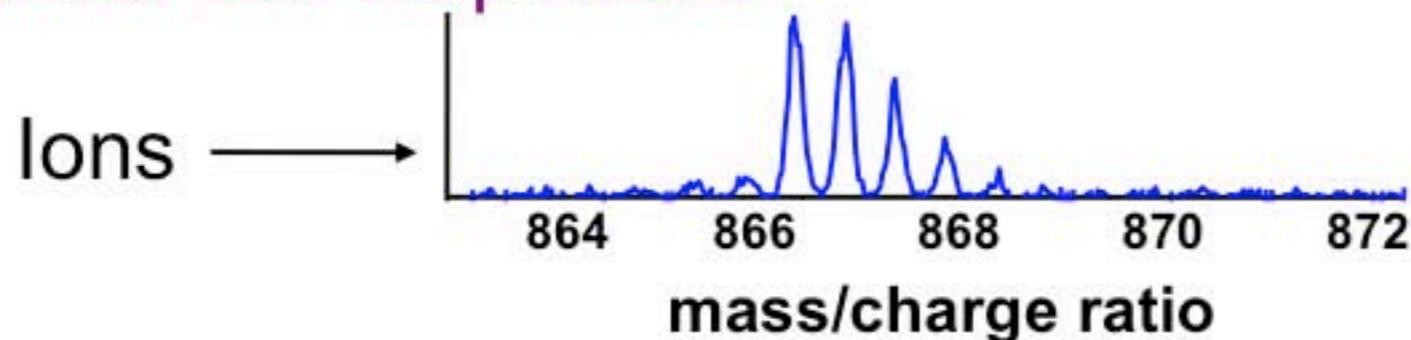
**>95% confidence**  
**>0% confidence**

**>50% confidence**  
no evidence

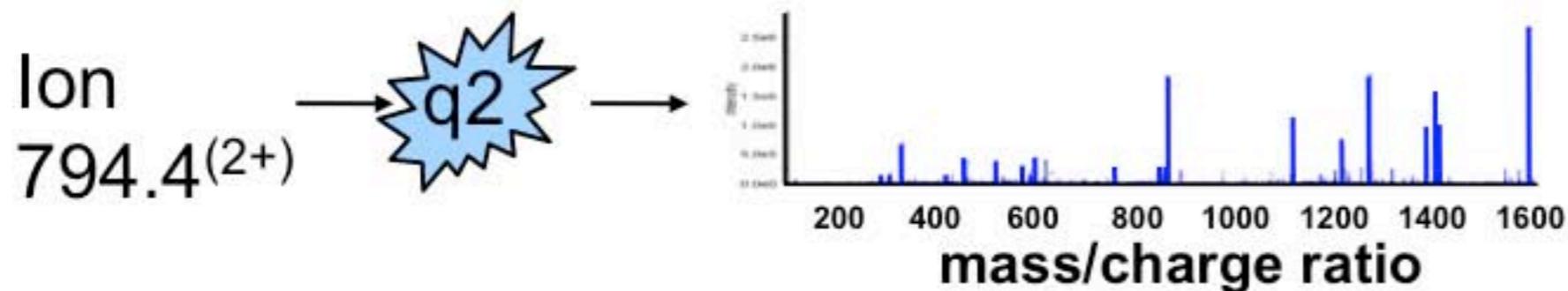
**How can we get  
absolute quantification  
of a specific protein in a  
complex sample?**

# ABI 4000 QTrap Can Operate Like a Normal Triple Quadrupole

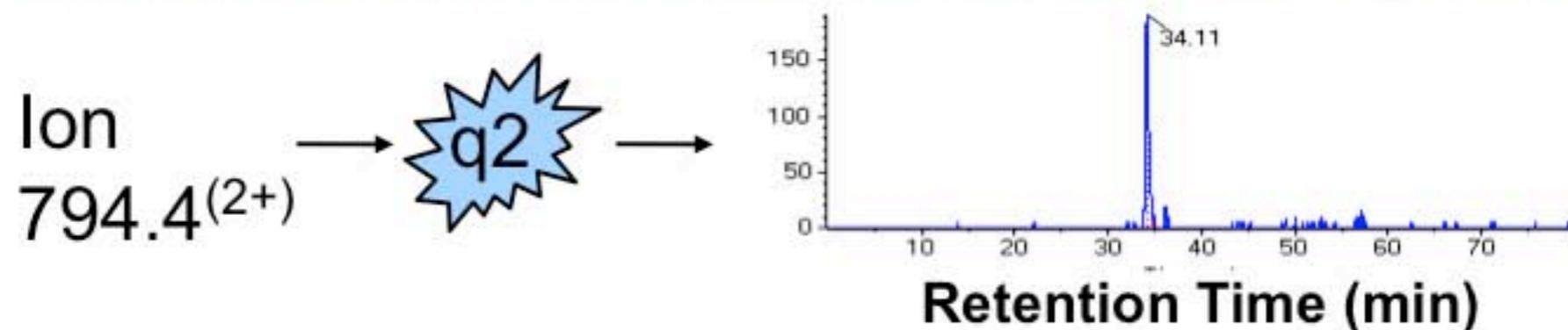
## Linear Ion Trap Mode



## Select a precursor, Fragment, Scan fragments



## Select a precursor, Fragment, Select specific $m/z$ on Q3

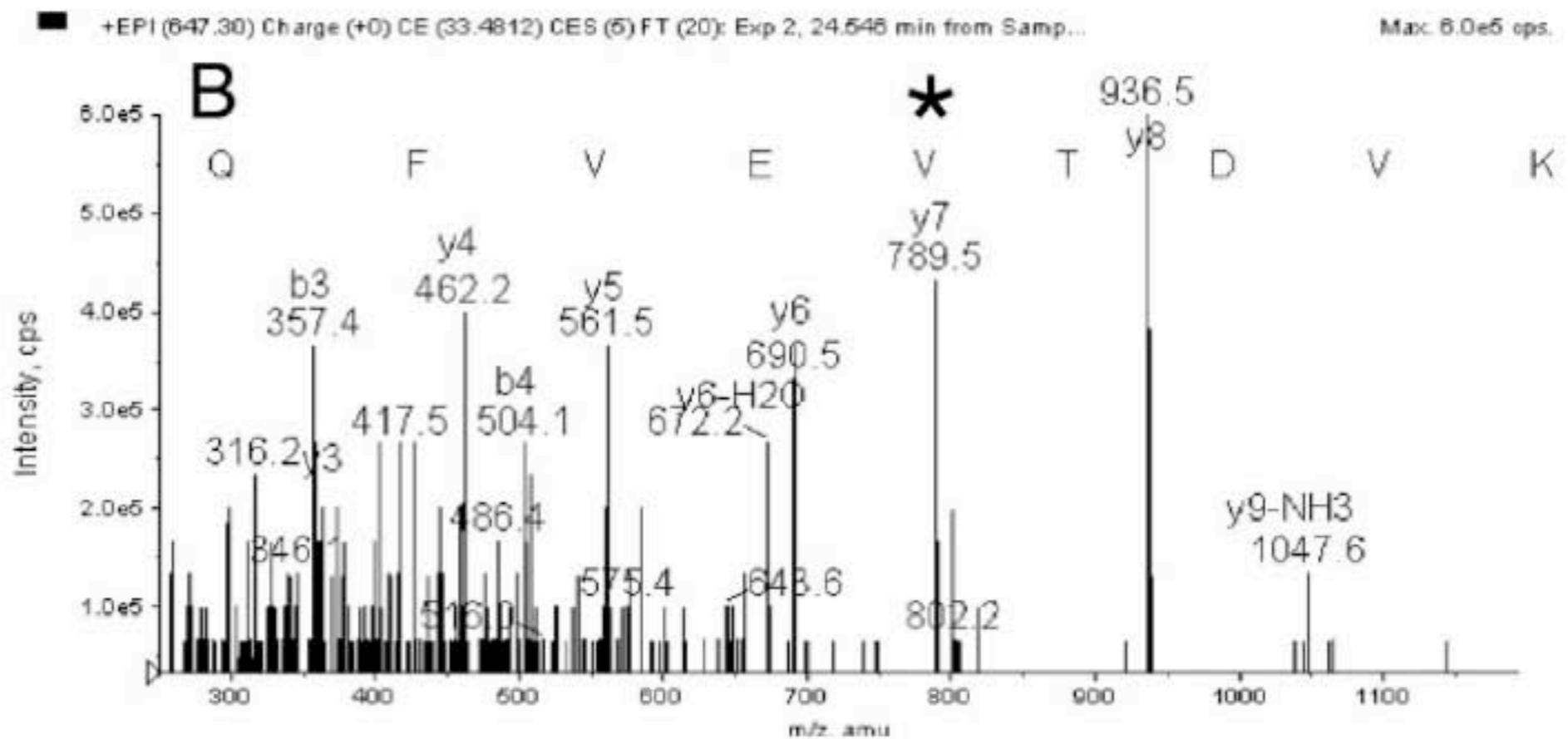
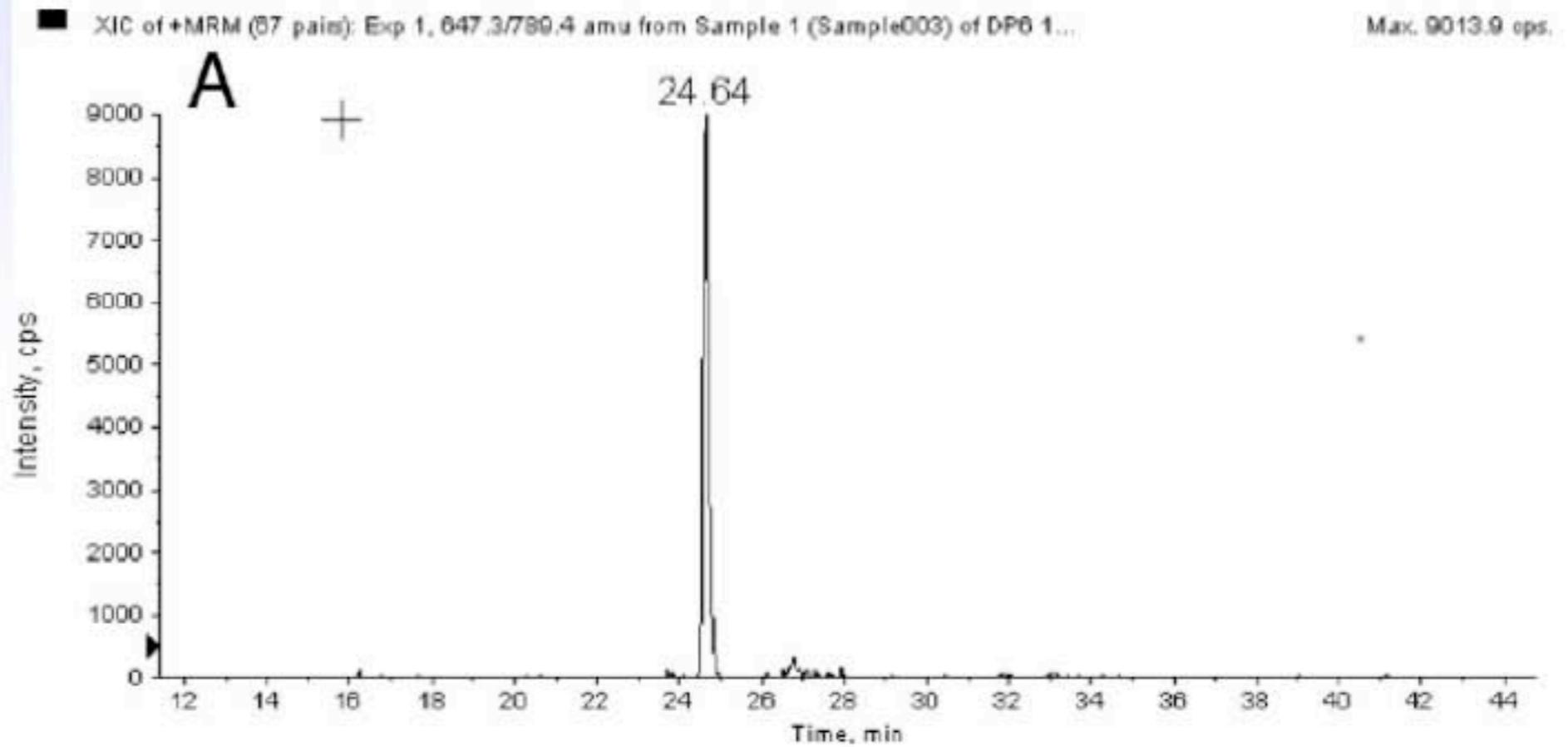


*Yields a chromatographic peak*

# Precedent in Medicine

Peptide of fibronectin, biomarker for preeclampsia (in urine) and emphysema (in plasma)

Anderson and Hunter, **2006**,  
*Mol. Cell. Proteomics* 573



Name for assay: Multiple reaction monitoring (MRM)

## **MRM Targets We Chose**

- 5 reductive dehalogenases, Hup, other oxidoreductases, RNA polymerase, ribosome subunit L7, S-Layer, GroEL
- Monitored 2 peptides per protein, 2 fragments per peptide

# Are these useful bioindicators?

## Trypsin “Digestion” of NCBI-nr

### Better Target Peptides:

WEGTPEENLLIMR: Would come from 19 TceA genes on NCBI-nr, all *Dehalococcoides* spp.

DEWWASENPIR: Would come from 19 TceA genes on NCBI-nr, all *Dehalococcoides* spp.

### Less-Specific Peptide:

FATSDLNDLYR: Comes from 3 *Dehalococcoides* genes and **1007** additional bacterial Rpo genes!

# Picking Transitions for Multiple Reaction Monitoring (MRM)

TceA peptide WEGT-PEENLMIR elutes around 57 min on nLC

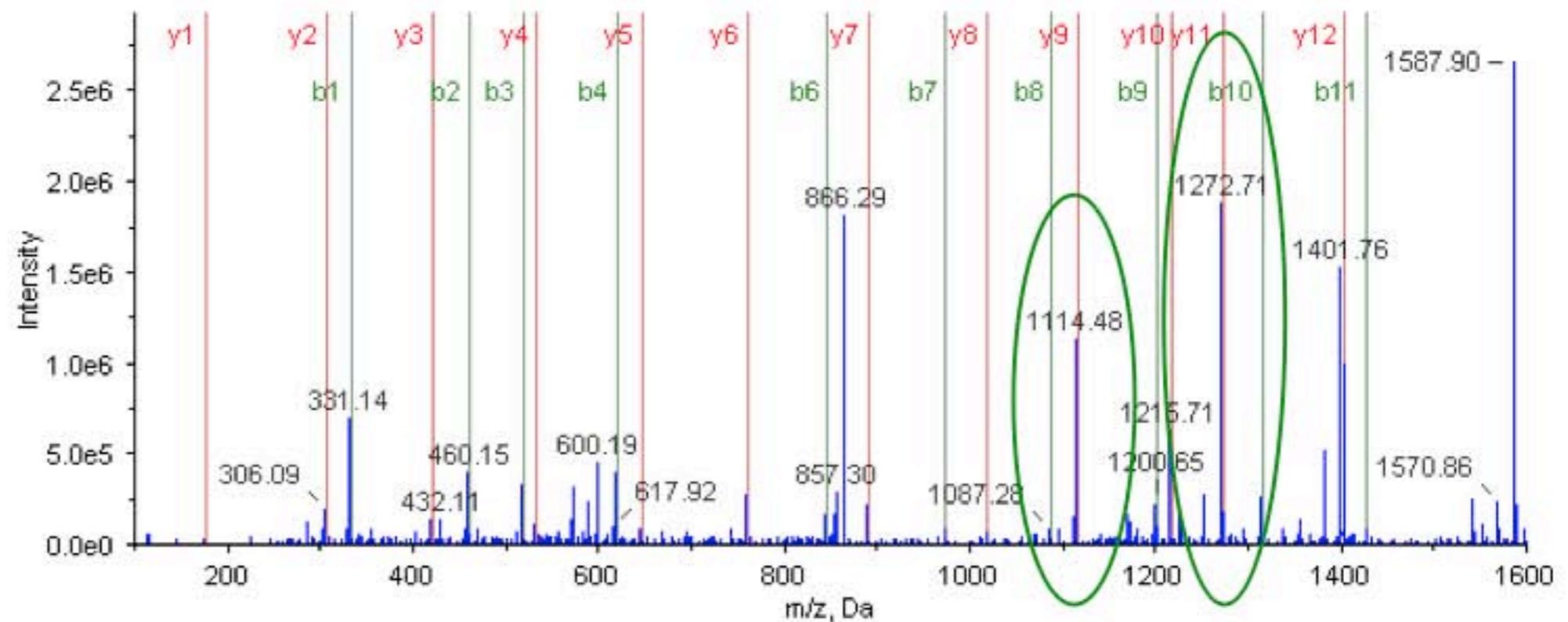
Ionization  
 →  
 (ESI)

Select on Q1:  
 Precursor ion 794.4<sup>(2+)</sup>

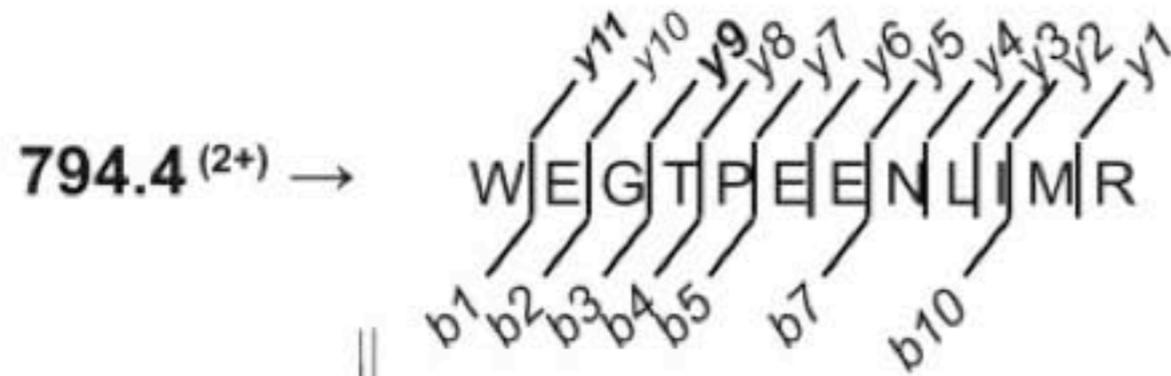
Fragmentation  
 in collision cell  
 (q2)

Pick representative fragments for Q3

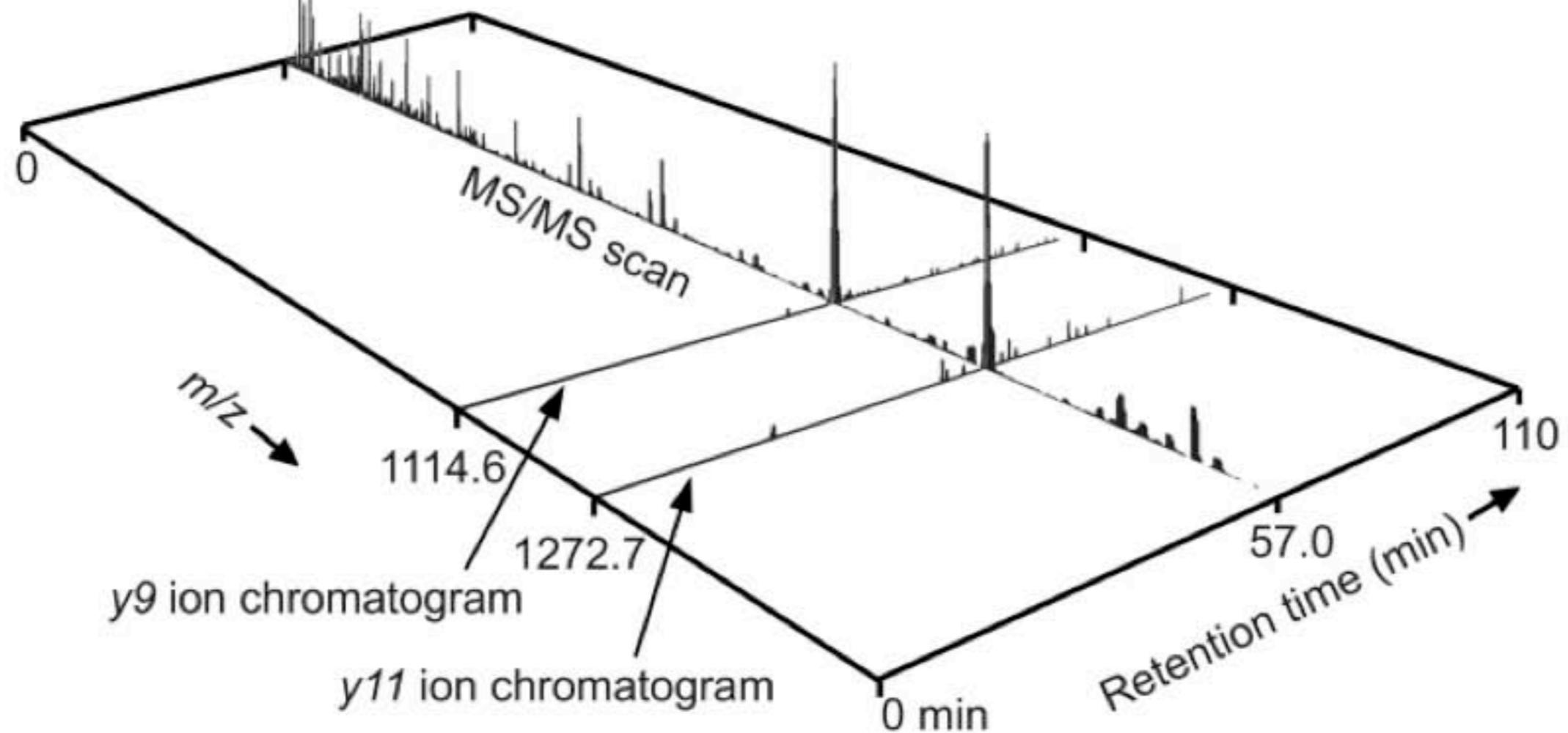
| Residue | b         | y         |
|---------|-----------|-----------|
| W       | 331.1887  | 1731.8856 |
| E       | 460.2312  | 1401.7042 |
| G       | 517.2527  | 1272.6616 |
| T       | 618.3004  | 1215.6402 |
| P       | 715.3532  | 1114.5925 |
| E       | 844.3957  | 1017.5397 |
| E       | 973.4383  | 888.4971  |
| N       | 1087.4813 | 759.4546  |
| L       | 1200.5653 | 645.4116  |
| L       | 1313.6494 | 532.3276  |
| I       | 1426.7335 | 419.2435  |
| M       | 1557.7739 | 306.1594  |
| R       | 1713.8751 | 175.1190  |



# Peptide Quantification *via* Multiple Reaction Monitoring

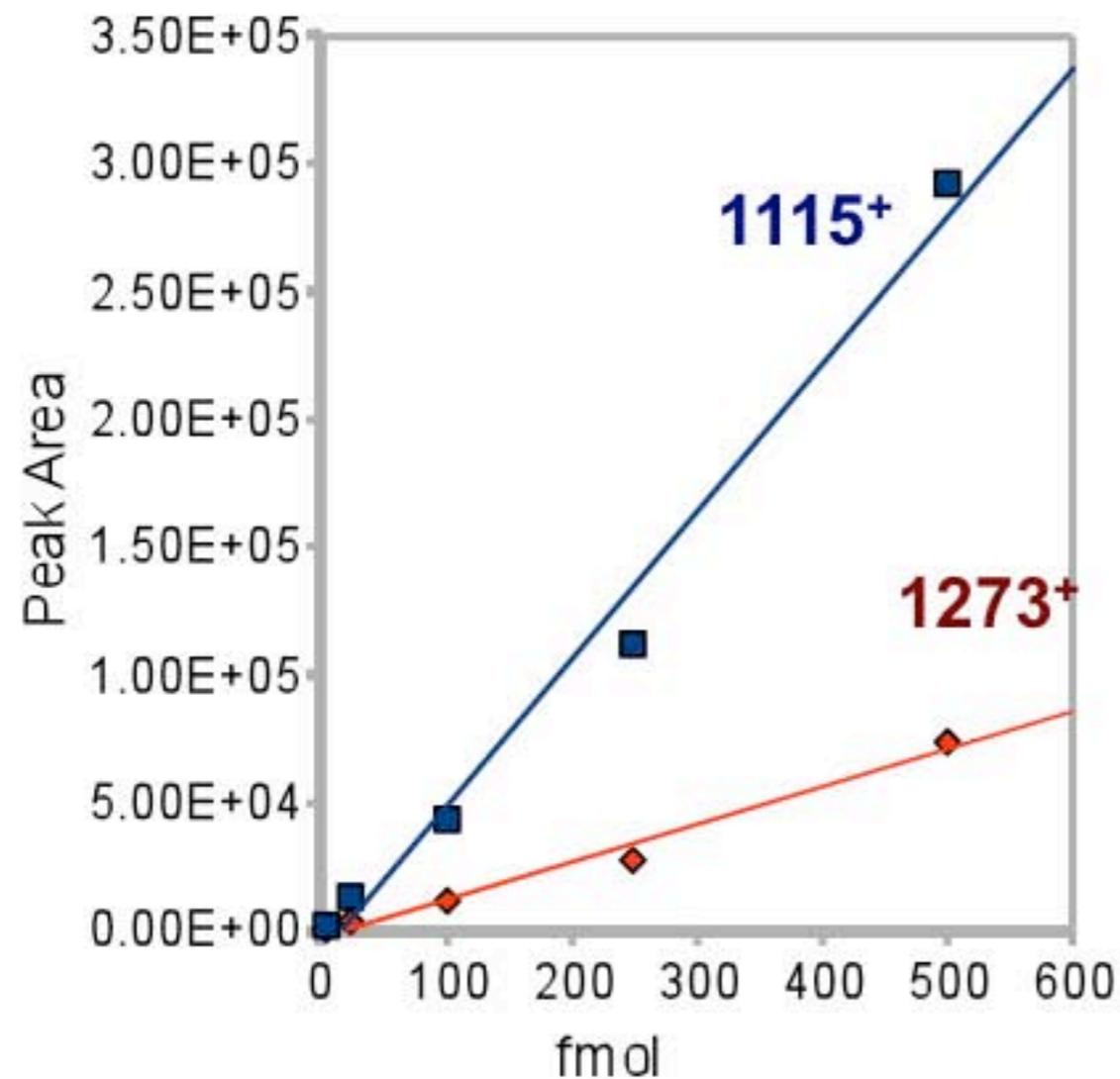


(TceA peptide)

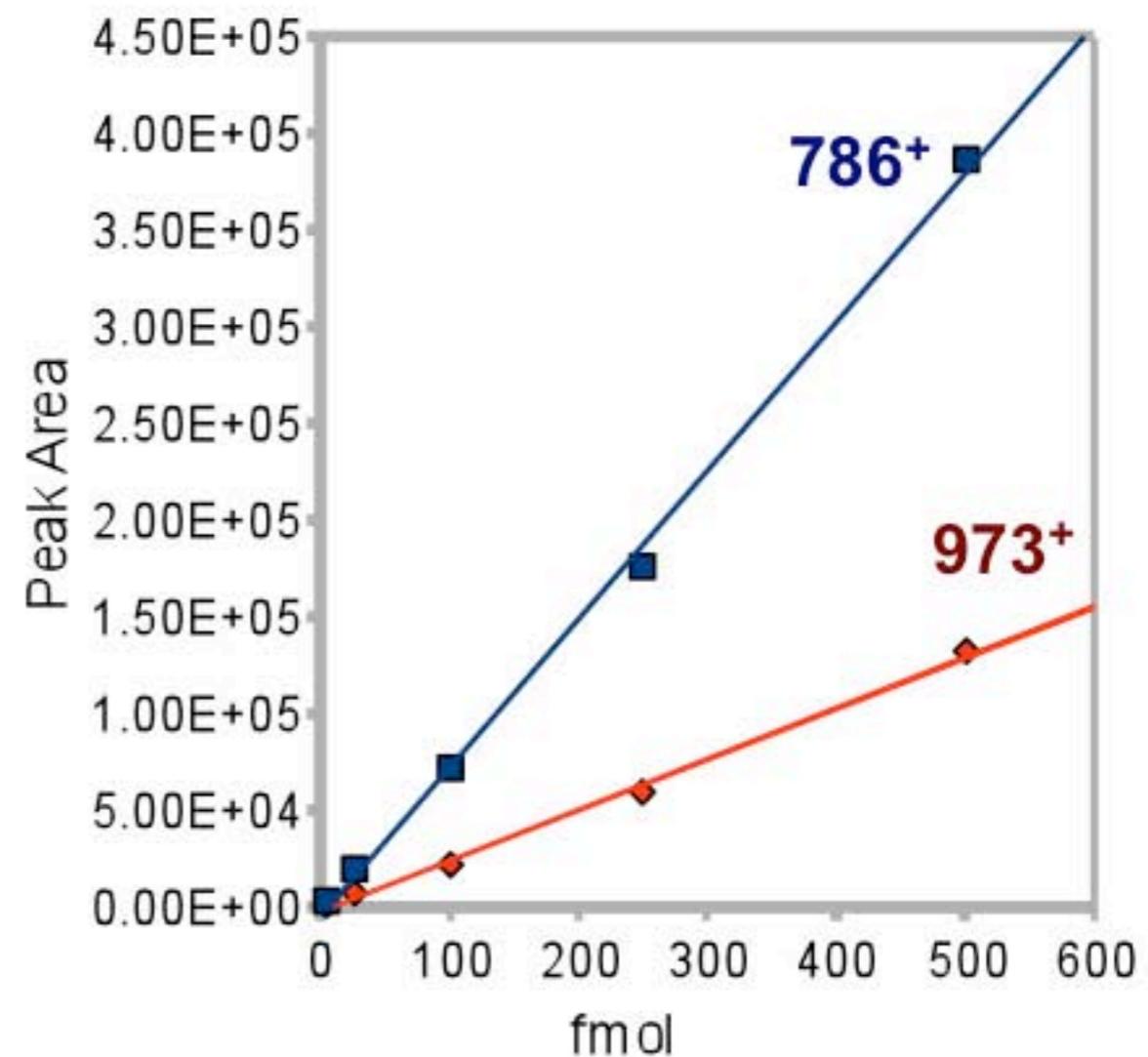


# Linear Response in a Background Sediment Digest

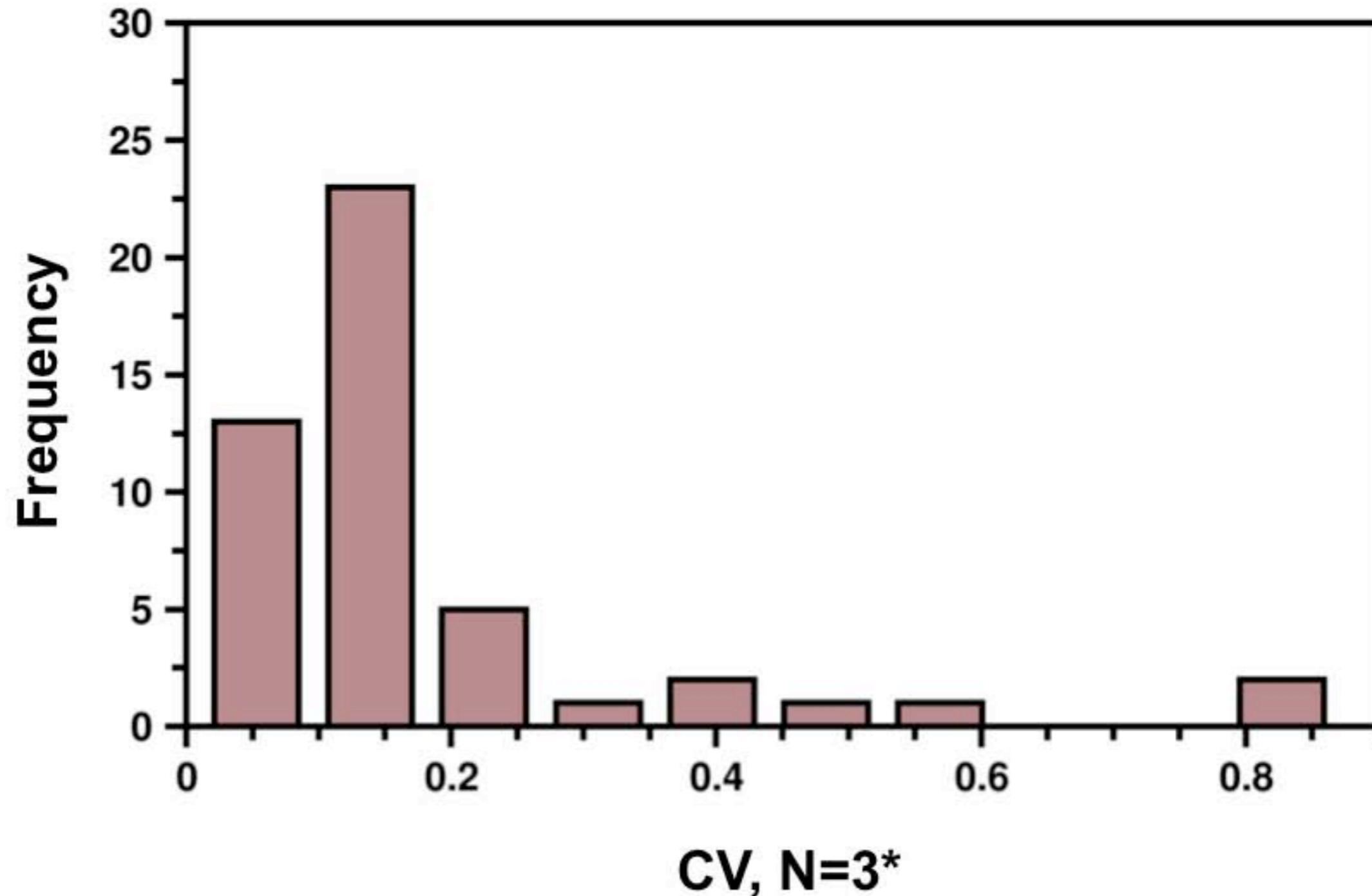
TceA: WEGTPEENLLIMR 794<sup>(2+)</sup> →



TceA: DEWWASENPIR 702<sup>(2+)</sup> →

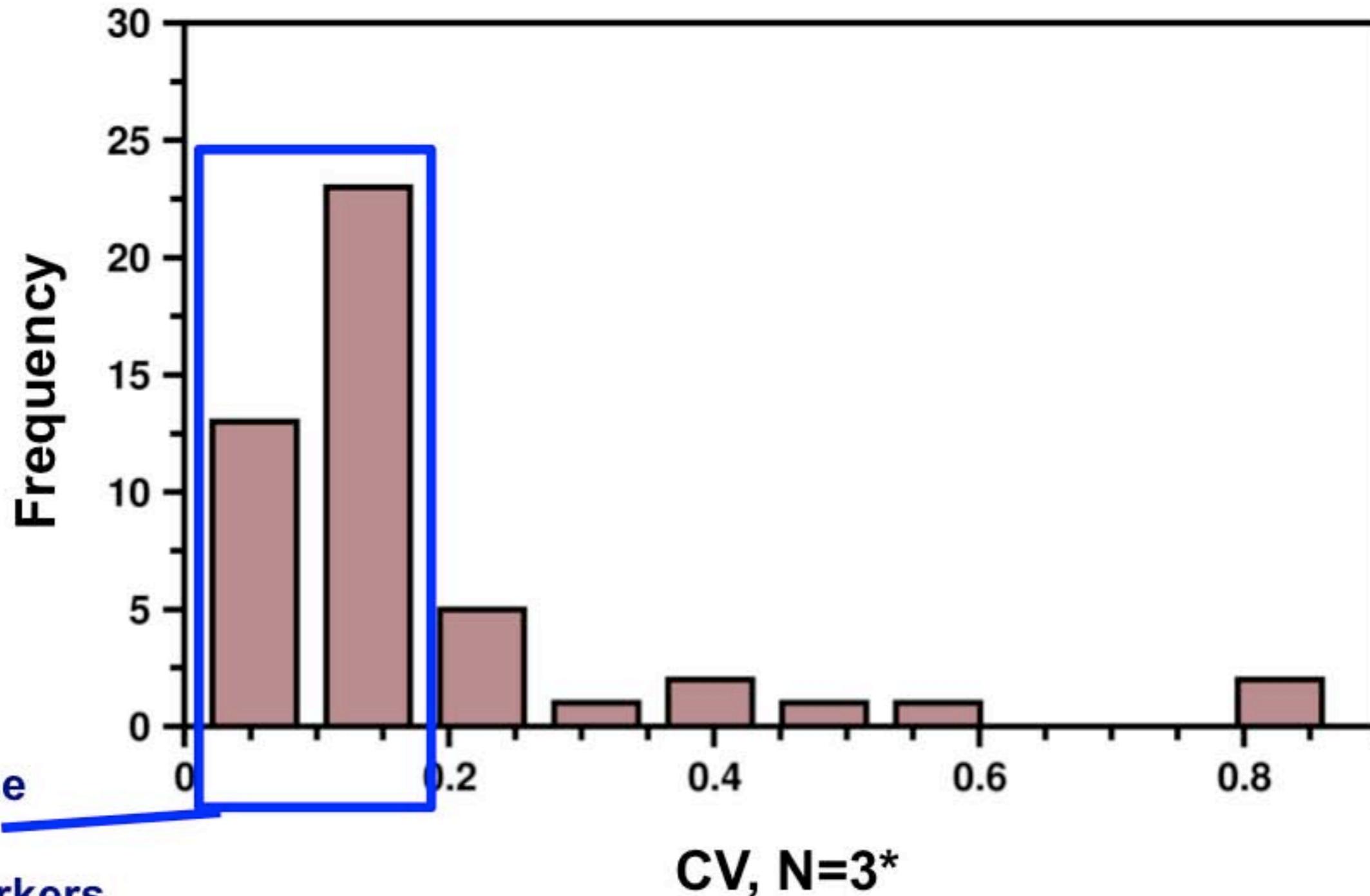


# Many Peptides Had Low Variability Over a 4 d Period



\*Independent analysis of 100 fmol standard on Day 1, Day 3 & Day 4

# Many peptides had low day-to-day variability



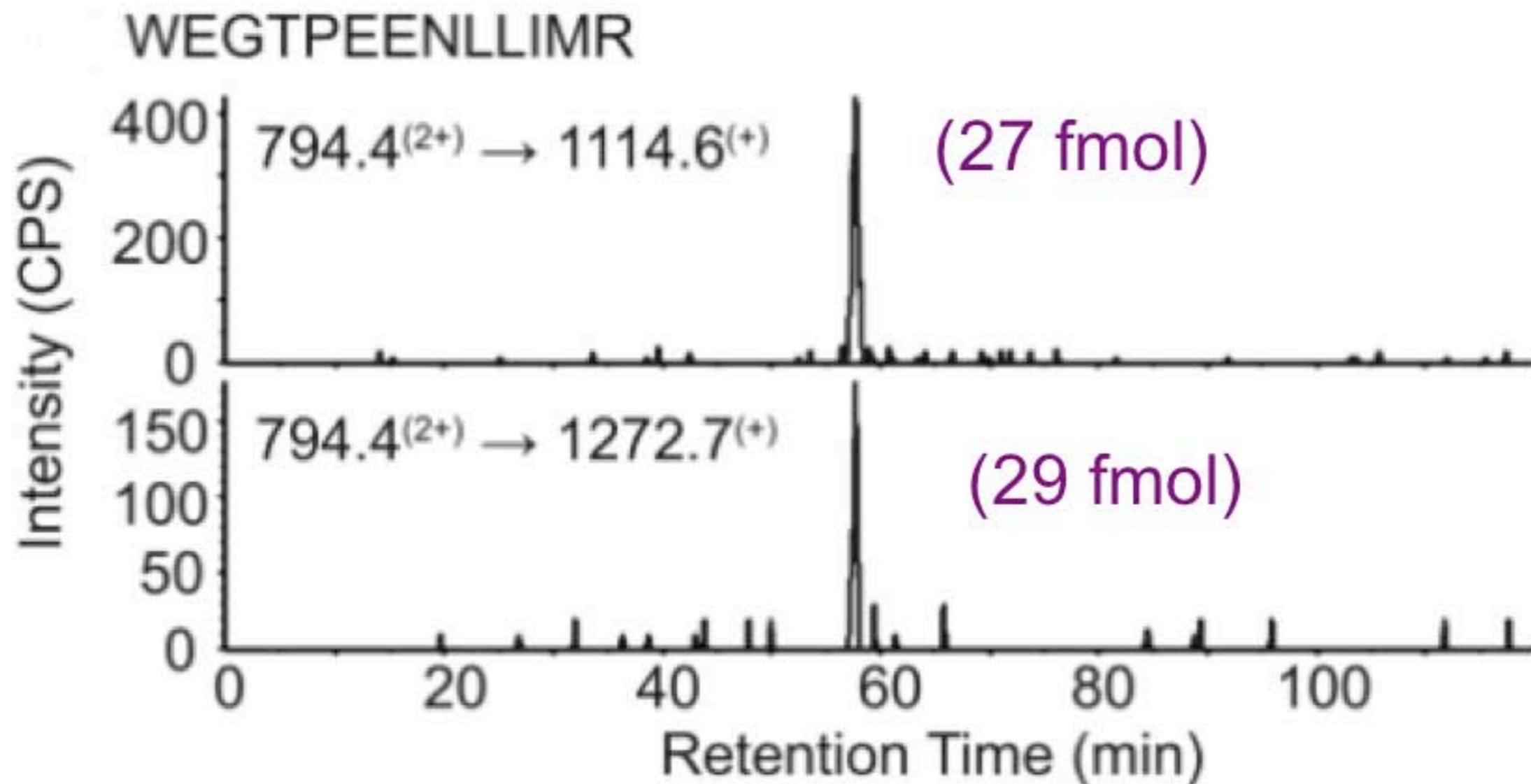
\*Independent analysis of 100 fmol standard on Day 1, Day 3 & Day 4

# **Most of the following data available in publication:**

Werner, Ptak, Rahm, Zheng and Richardson  
**2009** *Environmental Microbiology* 11:2687-2697

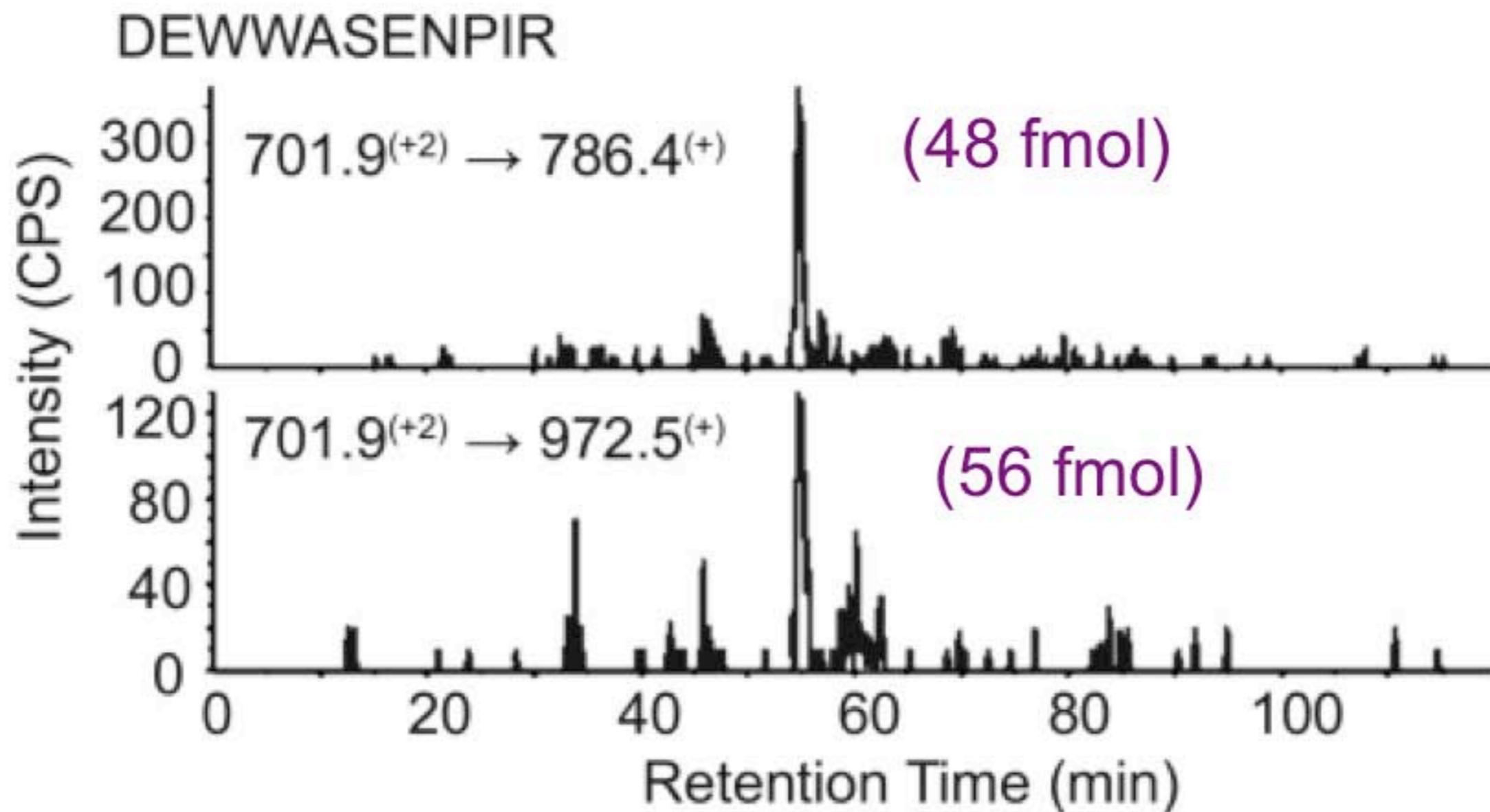
# TceA Enzyme ID and Quantification

## Gene Previously Thought to be Missing from Commercial Culture KB1™



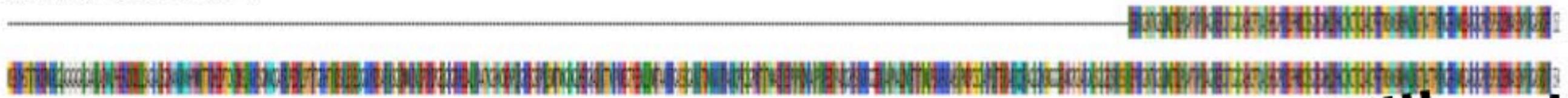
# TceA Enzyme ID and Quantification

## Gene Previously Thought to be Missing from Commercial Culture KB1™

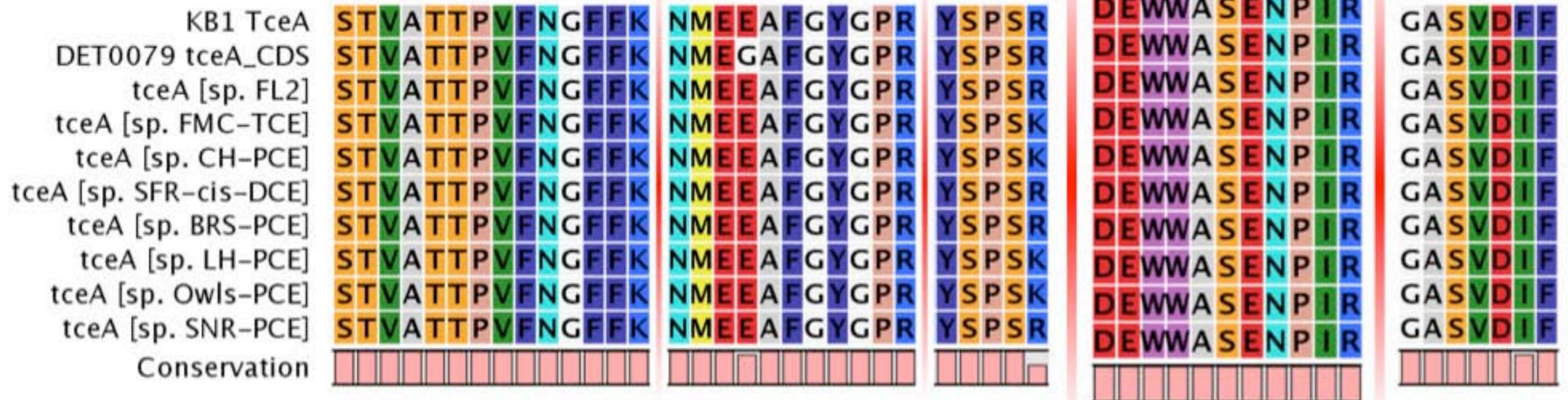


# Preliminary KB1 Metagenomic Sequencing Library

TceA from KB1

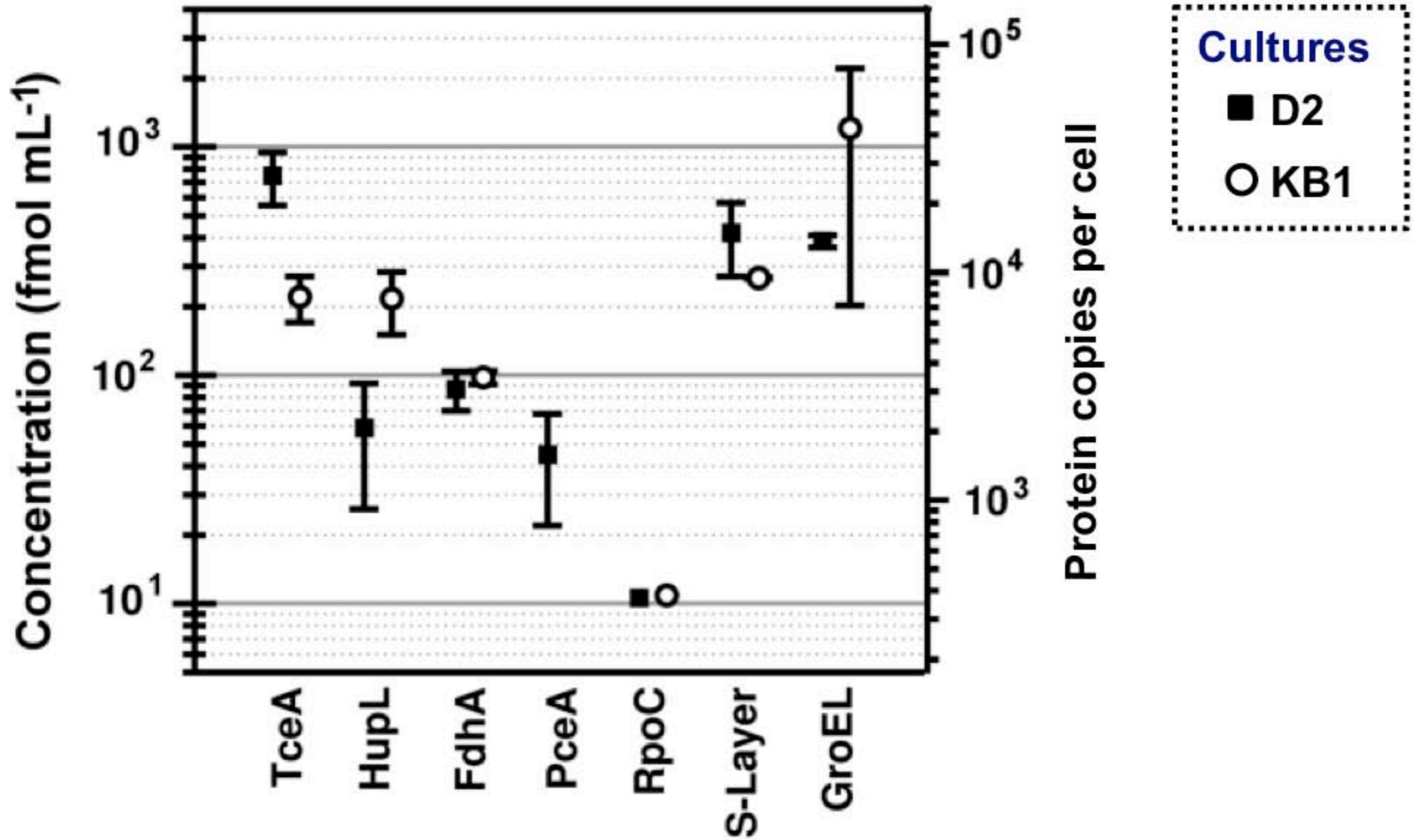


TceA from DET

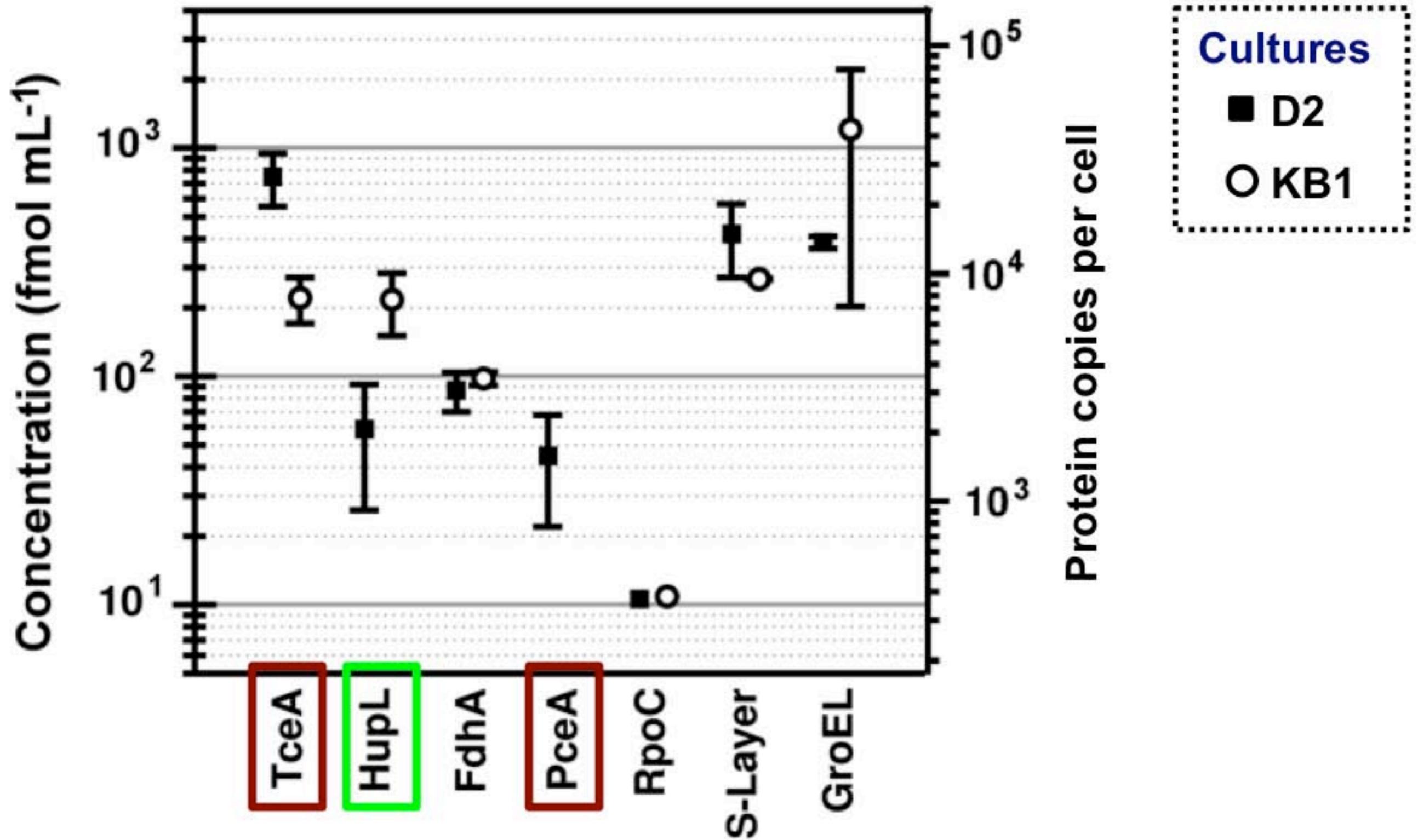


Preliminary KB1 gDNA fragment library from Elizabeth Edwards and Alison Waller at U. Toronto

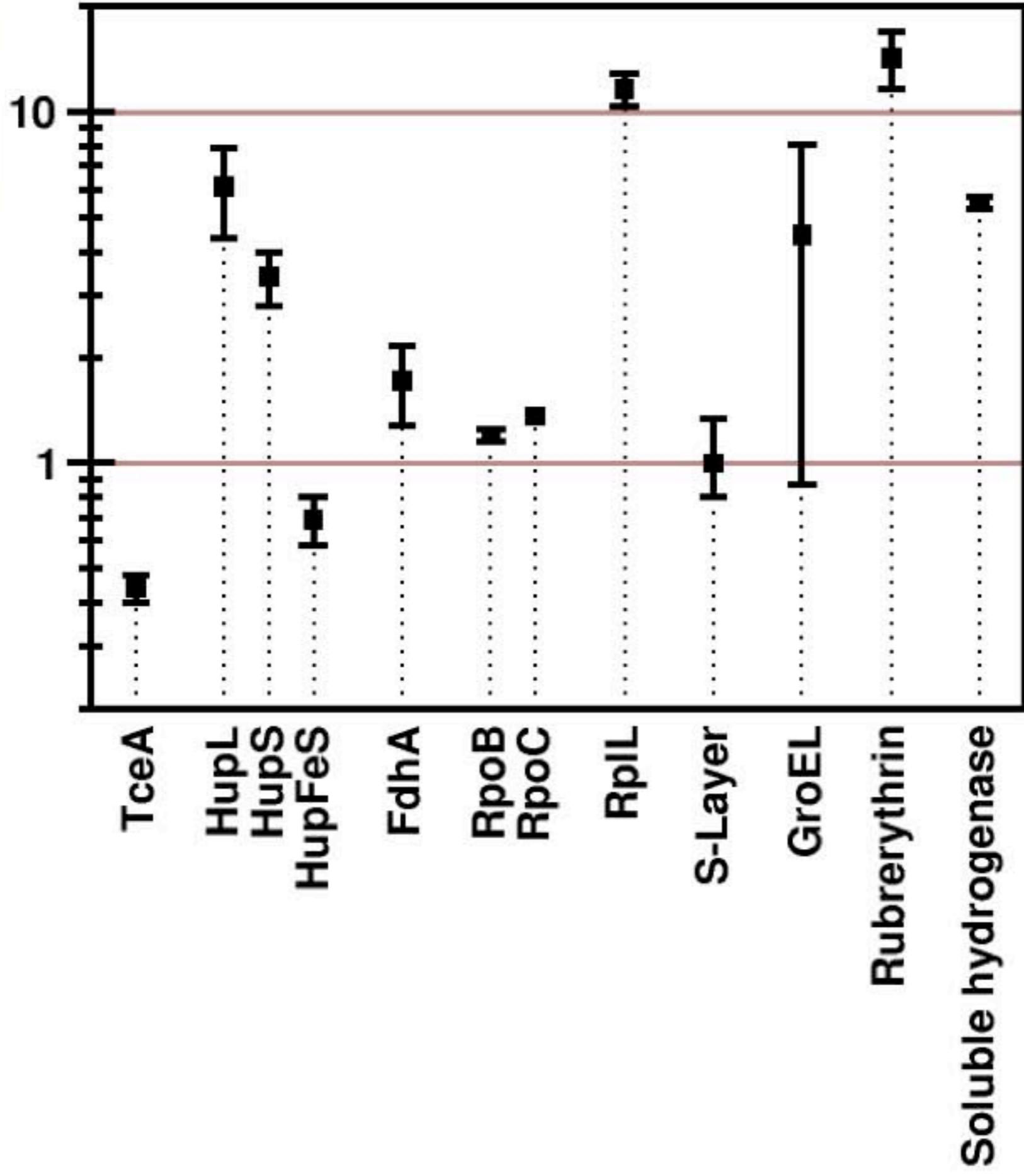
# Concentrations in Mixed Cultures



# Concentrations in Mixed Cultures



# S-layer normalized protein abundance ratio KB1/D2



# Challenges

- Quantification of RDases in Field Samples
  - 1-10 L of groundwater for active sites
  - Protein extraction protocol?
  - Need to define matrix effect on protein extraction and trypsin digestion
  - Trypsin digestion directly in sample/ acetonitrile? Peptides are easier to extract and to define extraction efficiency.

# Thank you

- Brian Rahm, Annie Rowe, Gretchen Heavner, Bob Morris, *Richardson Lab*

## **Funding**

Cornell Institute for Biotechnology, Innovations Grant  
Department of Defence  
National Science Foundation (award no. 0731169)