

Metaproteomics to investigate the impact of sampling-site biogeochemistry on structure and functionality of leaf-litter degrading microbial communities

Thomas Schneider, Katharina Keiblinger, Bertran Gerrits, Emanuel Schmid, Leo Eberl, Sophie Zechmeister-Boltenstern, Kathrin Riedel



PD Dr. Kathrin Riedel
Microbial Proteomics
Institute of Plant Biology
University of Zurich
Winterthurerstrasse 190
CH-8057 Zürich





MICROBIAL PROTEOMICS GROUP ZURICH



OTHER RESEARCH INTERESTS

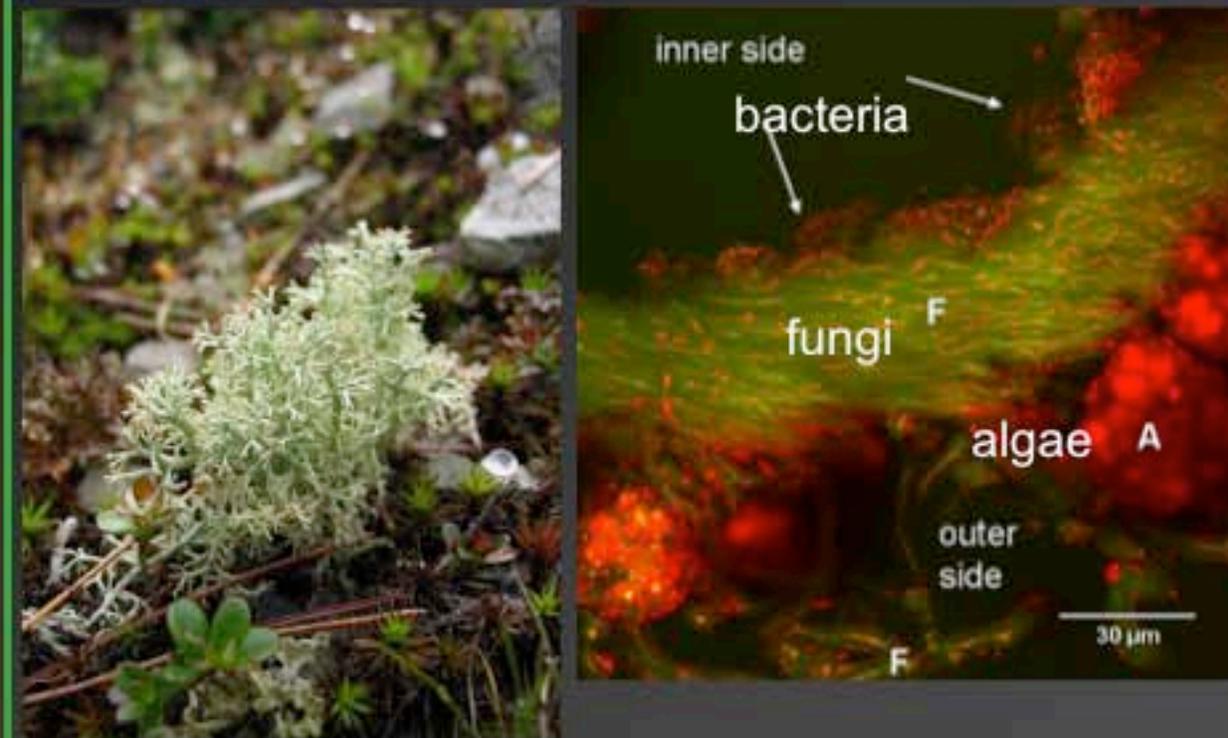
Global Warming & Microbial Biofilms in Rivers



Which effects have drying and re-wetting on structure and function of microbial biofilms in European rivers?

Future cooperation with universities in Vienna (T. Battin), Rome (S. Fazio) and Giessen (J. Marxsen)

Microbial Symbiosis in Lichens



How are lichens protected against desiccation, UV-light, and coldness?
What is the molecular basis of the symbiosis?

Cooperation with the TU Graz (G. Berg) and the University of Graz (M. Grube)

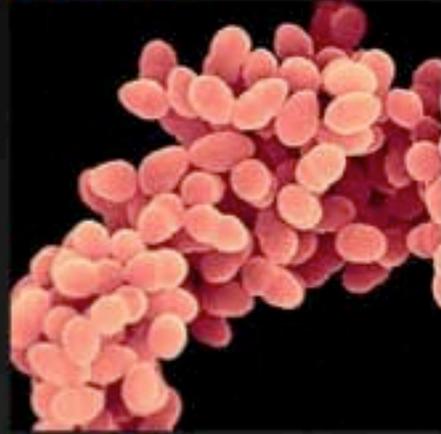


OTHER RESEARCH INTERESTS

PATHOGENS



*Pseudomonas
aeruginosa*



*Staphylococcus
aureus*

in presence / absence of
drug candidates

INFECTION MODELS



*Caenorhabditis
elegans*



Murine model

COMPARATIVE QUANTITATIVE PROTEOME ANALYSES

of infected hosts in the absence and
presence of drug candidates

global changes in protein
expression in both the host and the
pathogen in response to the drug
treatment

„METAPROTEOMICS“

Advantage: genomic sequence of
hosts and pathogens is available.



MOTIVATION & HYPOTHESES

- **Yearly litter production:**

 - ~ 10^{11} t (Drymass)

 - Large pool of organic carbon, nitrogen & phosphorous

- **Global mean steady state turnover time:**

 - ~ 1.4 to 3.4 years



HYPOTHESES

- ⇒ in most aerobic habitats fungi are the main degraders of litter
- ⇒ in anaerobic habitats or in environmental niches with increased temperature, bacteria contribute to litter decomposition
- ⇒ community structure and function is influenced by leaf-litter biogeochemistry



GOALS & EXPERIMENTAL DESIGN



**LEAF LITTER FROM
DIFFERENT SAMPLING
SITES
(February & May)**

**Protein separation
and identification
1D-PAGE LC-MS/MS
Orbitrap**

Affiliation of proteins to different phylogenetic & functional groups



Bacteria



Fungi



Protozoa



Plants



Insects

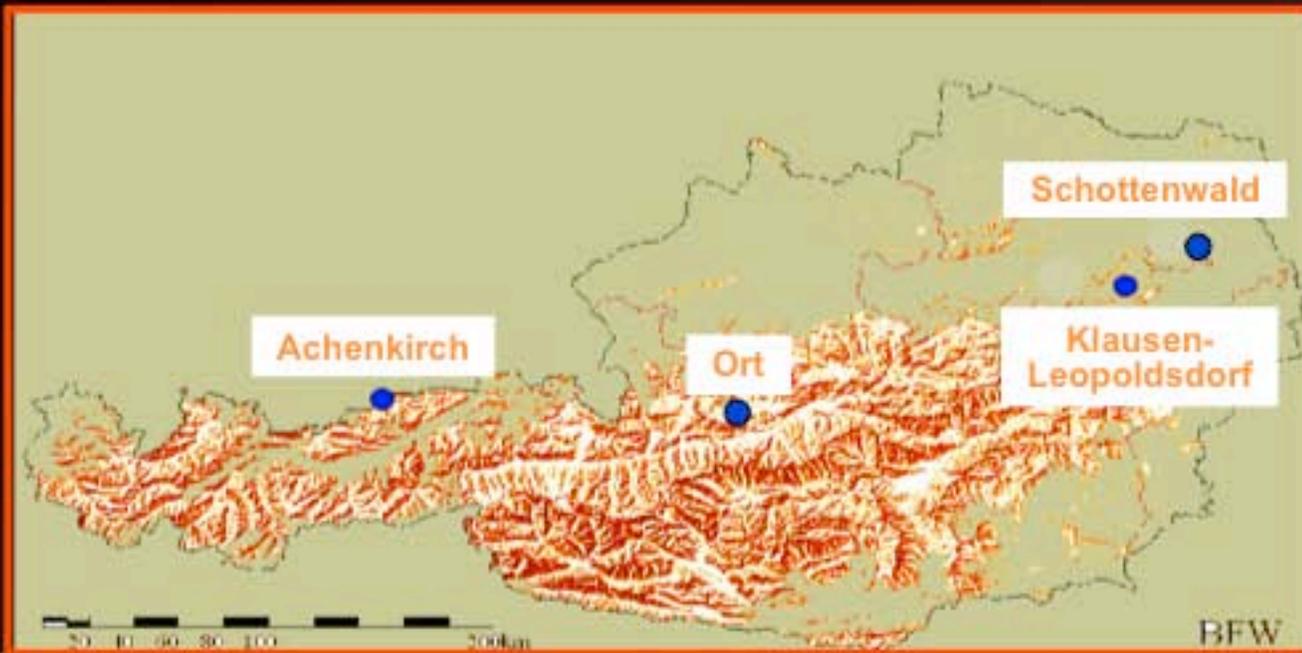


Animals

Contribution of metabolically active organisms to the respective habitat



LITTER BIOGEOCHEMISTRY (FEBRUARY)



Sophie Zechmeister-Boltenstern
Katharina Keiblinger
BFW - Bundesamt für Wald, Vienna

	Ort	Achenkirch	Klausen-Leopoldsdorf	Schottenwald	Schottenwald
Water content [%]	57	73	79	77	68
C [g/kg]	472	502	468	455	470
N [g/kg]	8.2	12.4	10.0	12.5	10.0
C:N Ratio	57.8	40.3	46.8	36.4	47.0
P [g/kg]	0.35	0.03	0.04	0.64	0.22
C:P Ratio	1361	1439	1160	714	2175
Mn [ppm]	1041	88	832	1493	1306
	Beech litter			Oak litter	



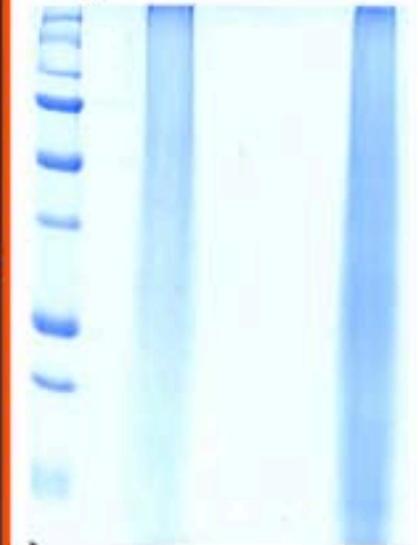
1D-SDS-PAGE-LC-MS/MS



Leaf litter from different sampling sites and seasons

- Protein extraction (SDS-cont. buffer)
- Sonication / Centrifugation
- Concentration (Speedvac)

Prefractionation
by 1-D PAGE



MS and MS/MS data

- Assignment of proteins to functional & phylogenetic groups
- Semi-quantitative analysis by spectral counting

Database: UniRef100

in total assignment of about
~ 2000 proteins/protein clusters

Trypsin
Digestion

Peptide mixture



Peptide Separation & Analysis

LC-MS/MS

(MS LTQ OrbiTrap)



PROTEIN ASSIGNMENT & QUANTIFICATION

MS & MS/MS data

Database search (Mascot, X-tandem)
Data filtering and sorting (Scaffold:
95% peptide probability, 1 unique peptide)



**Clusters of proteins identified
by the same set of peptides**

Quality control: cluster homology
(> 50% homology)



**Validated clusters of proteins
identified by the same
set of peptides**

Assignment to phylogenetic groups
according to UniPROT NEWT taxonomy
Quality control - cluster consistency



**Validated clusters assigned to
a certain phylogenetic group**

Assignment to functional groups according
to KEGG, KOG, COG, and SWISS PROT
Quality control - cluster consistency



**Validated clusters assigned to
a certain functional group**

Quantification by counting of unique spectra assigned to taxonomic or functional groups



Emanuel Schmid

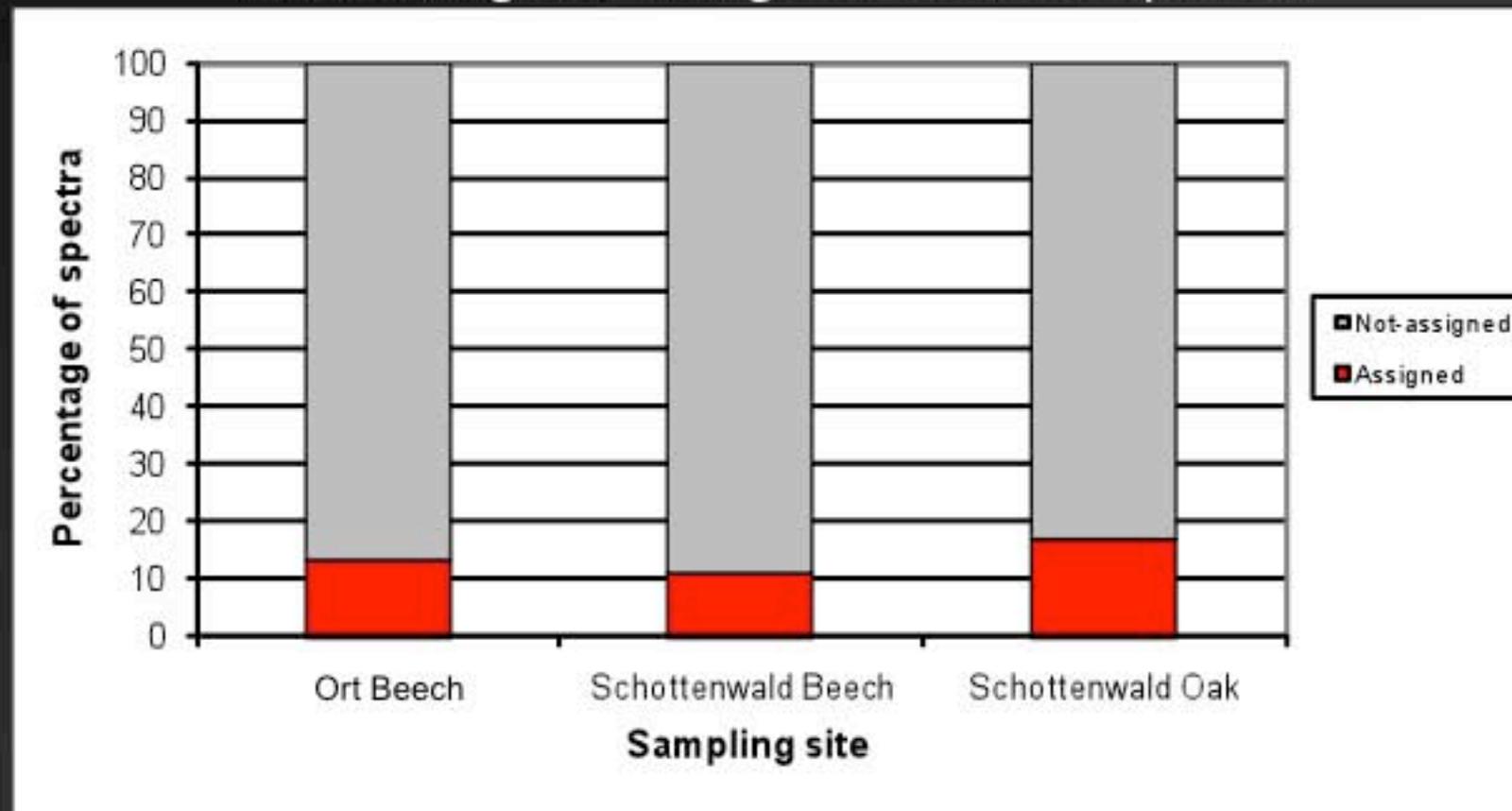
**Data analysis pipeline
using
PERL based scripts**



DRAWBACKS & CHALLENGES

- number of acquired MS/MS-spectra: ~300,000
- number of proteins: ~1200

Percentage of assigned MS-MS spectra

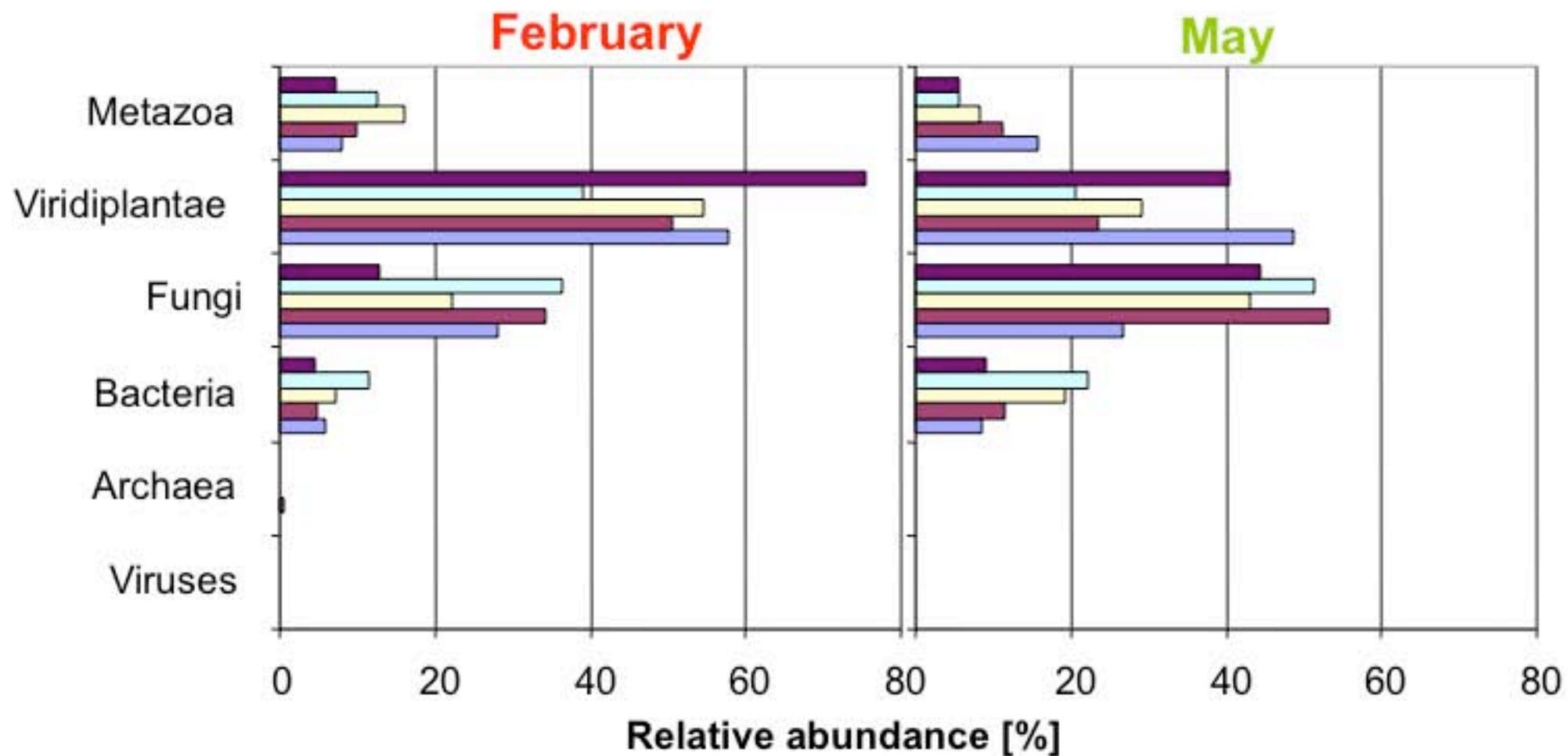


⇒ 90% of the spectra do not match to any protein in the reference database

⇒ database has to be improved (metagenomics & metatranscriptomics sequence data)



COMMUNITY STRUCTURE - OVERVIEW



- Schottenwald oak (422/1347)
- Schottenwald beech (649/1609)
- Klausen Leopoldsdorf beech (591/1375)
- Ort beech (673/1839)
- Achenkirch beech (955/2292)

Quantification is based on the numbers of unique spectra

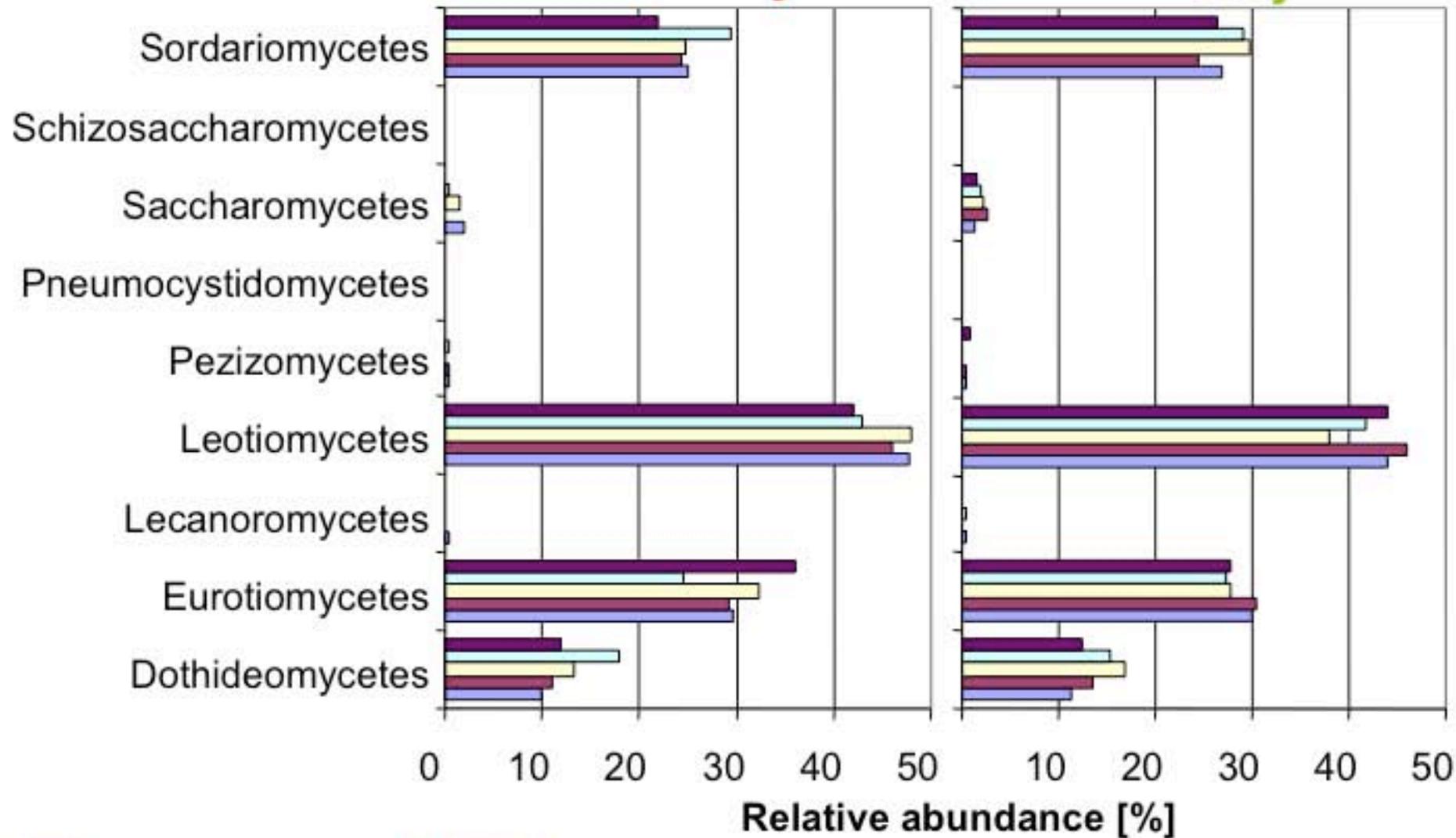


COMMUNITY STRUCTURE - FUNGI

February

May

Ascomycota

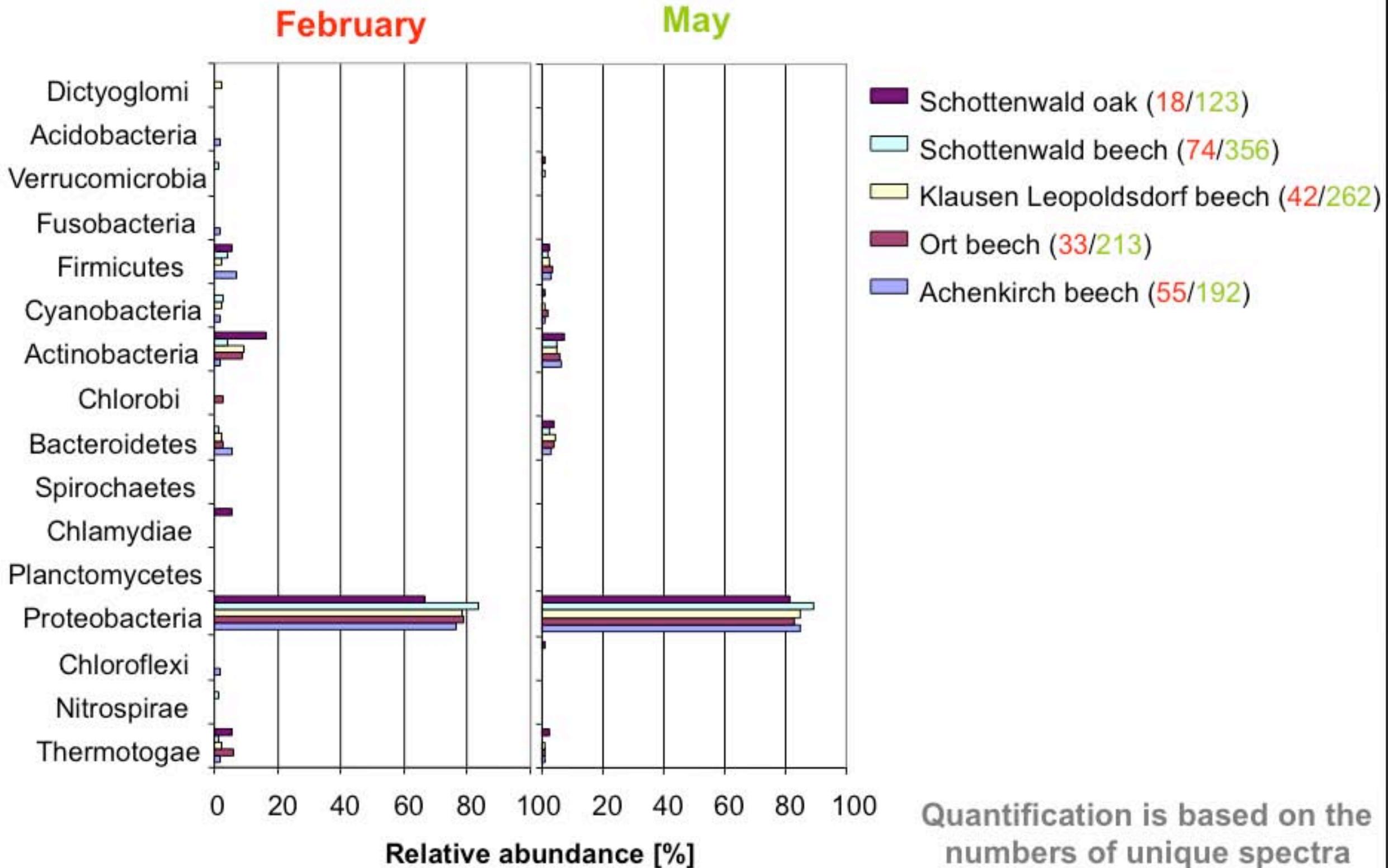


- Schottenwald oak (50/524)
- Schottenwald beech (228/713)
- Klausen Leopoldsdorf beech (121/512)
- Ort beech (226/801)
- Achenkirch beech (259/505)

Quantification is based on the numbers of unique spectra

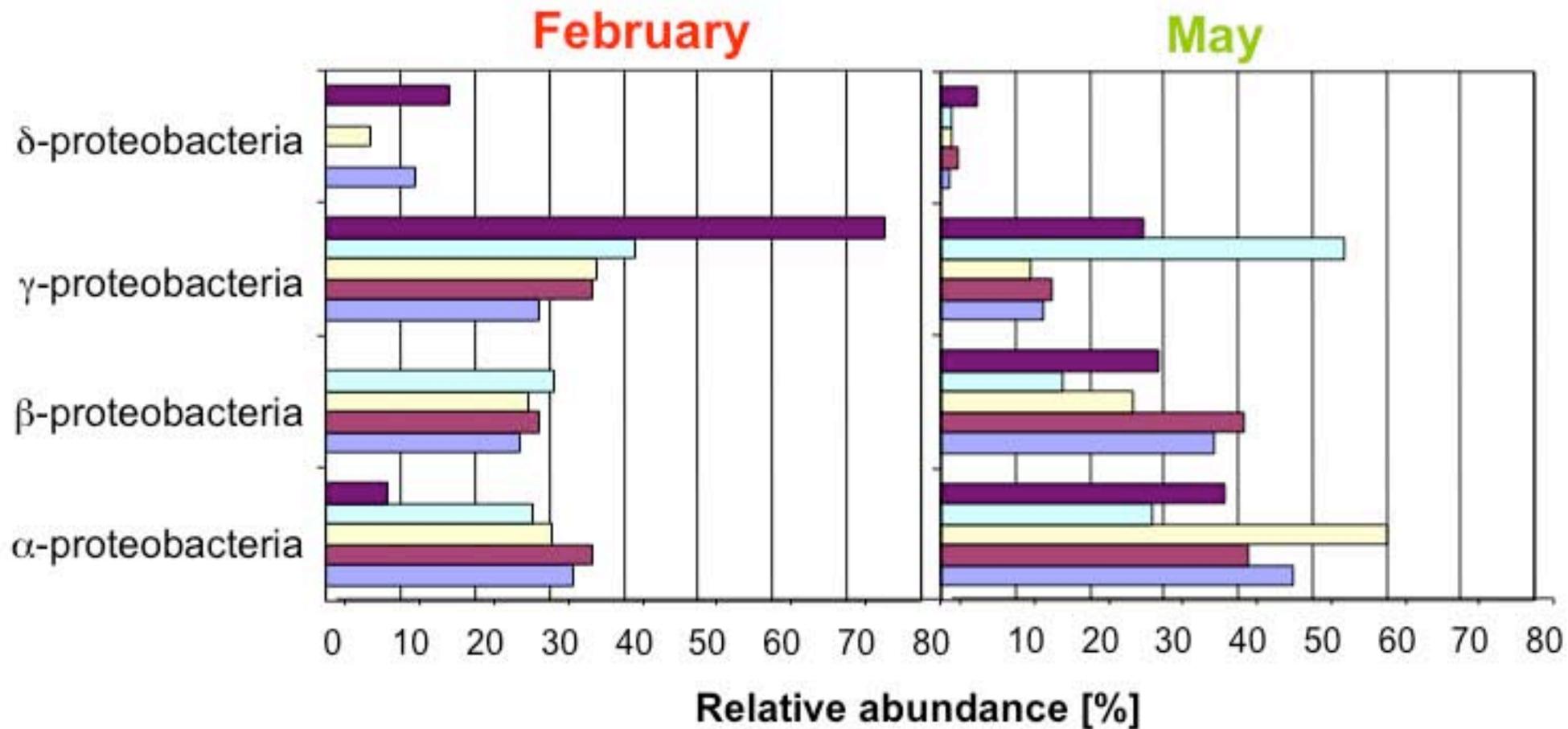


COMMUNITY STRUCTURE - BACTERIA





COMMUNITY STRUCTURE - BACTERIA



- Schottenwald oak (12/102)
- Schottenwald beech (65/322)
- Klausen Leopoldsdorf beech (33/223)
- Ort beech (28/179)
- Achenkirch beech (42/166)

Quantification is based on the numbers of unique spectra

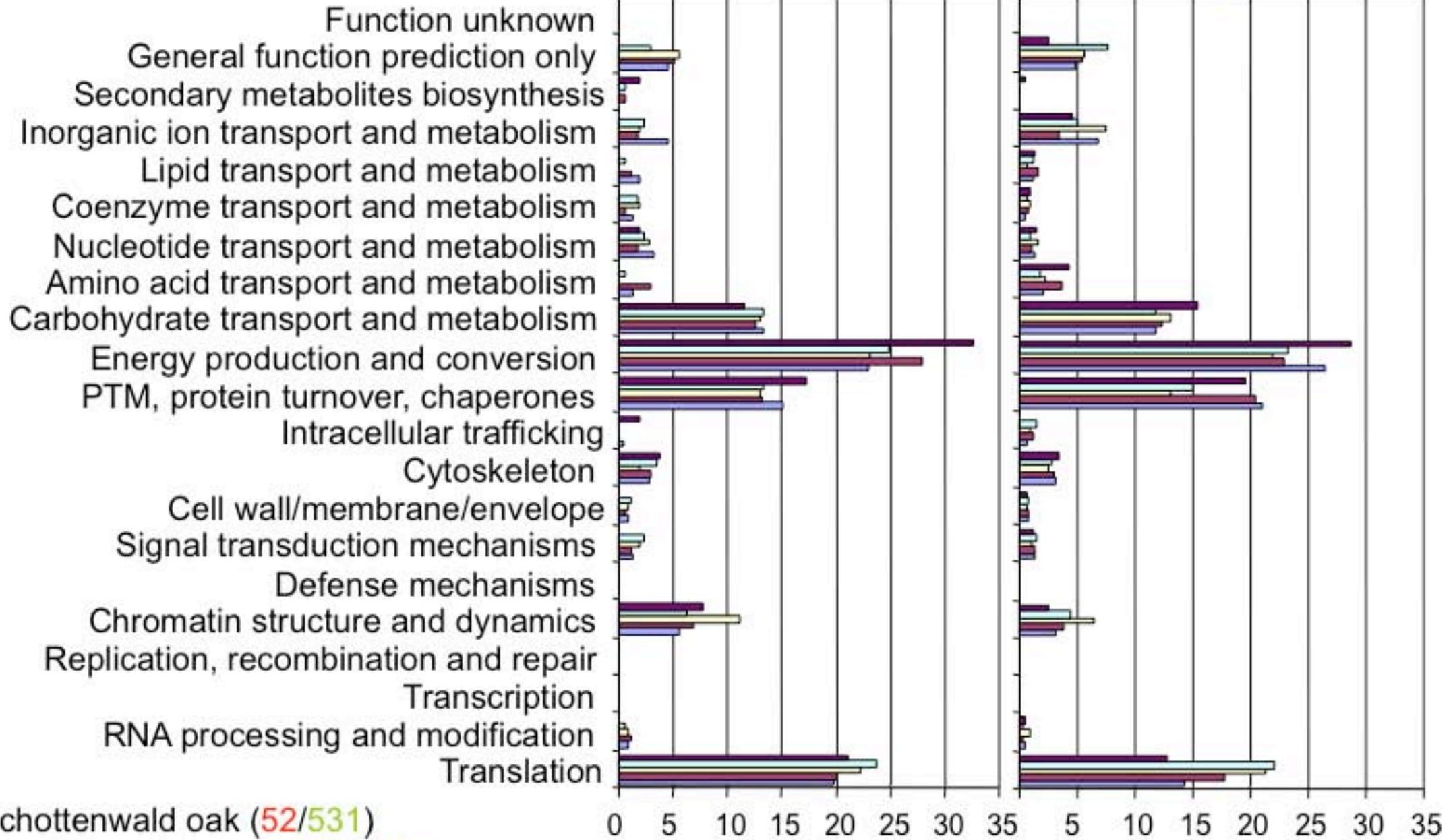


COMMUNITY FUNCTION - FUNGI

KOG classification

February

May



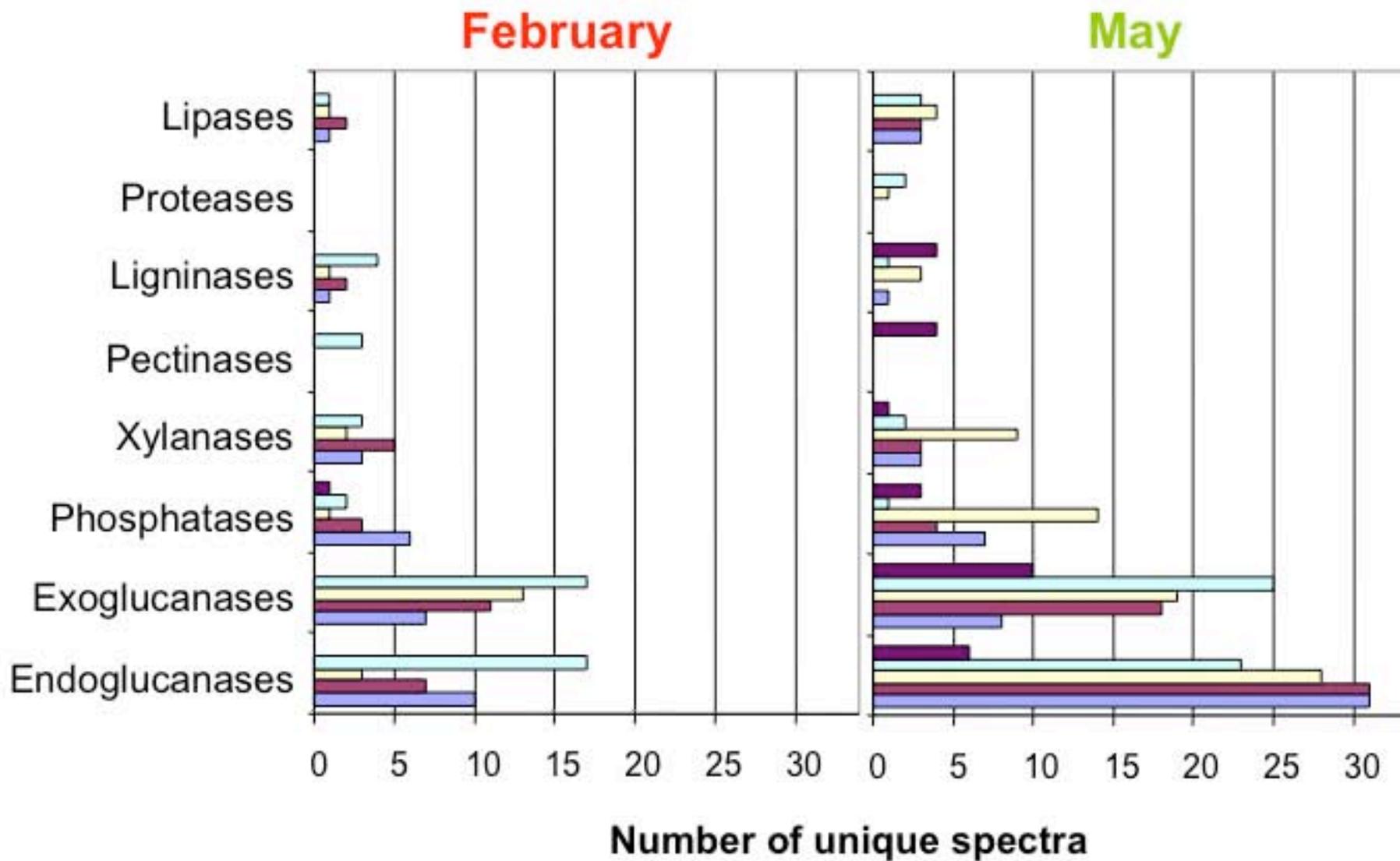
- Schottenwald oak (52/531)
- Schottenwald beech (173/708)
- Klausen Leopoldsdorf beech (108/483)
- Ort beech (175/859)
- Achenkirch beech (218/518)

Relative abundance
[%]

Quantification is based on the
numbers of unique spectra



COMMUNITY FUNCTION – FUNGAL EXOENZYMES



Hydrolytic enzymes are only of fungal origin

- Schottenwald oak
- Schottenwald beech
- Klausen Leopoldsdorf beech
- Ort beech
- Achenkirch beech

Quantification is based on the numbers of unique spectra



IMPACT OF LITTER BIOGEOCHEMISTRY?

Sampling site		N	P	Mn	Plant Proteins	Fungal Proteins	Bacteria Proteins	Fungal Exoenzymes
Litter type								
Achenkirch	Feb	+++	+	+	+++	++	+	++
Beech	May	++	+	+	+++	++	++	+++
Klausen-Ld.	Feb	++	+	++	++	++	+	+
Beech	May	+	+	++	+	+++	+++	+++
Ort	Feb	+	++	++	++	+++	+	++
Beech	May	++	++	++	+	+++	++	++
Schottenwald	Feb	+++	+++	+++	+	+++	++	+++
Beech	May	+++	+++	+++	+	+++	+++	+++
Schottenwald	Feb	+++	++	+++	+++	+	+	(+)
Oak	May	++	++	++	++	++	++	++

+, ++, and +++ refer to relative abundances between different sampling & litter types



DATA VALIDATION

ENZYMATIC ACTIVITY MEASUREMENTS

Sampling Site Litter Type	Cellulase (Units / ml)		Protease (Units / ml)		Xylanase (Units / ml)	
	Feb	May	Feb	May	Feb	May
	Achenk. Beech	0.007	0.01	0.12	0.1	0.025
Klau.-L. Beech	0.008	0.01	0.12	0.13	0.032	0.039
Ort Beech	0.013	0.018	0.18	0.2	0.04	0.06
Sch.W. Beech	0.009	0.016	0.25	0.3	0.035	0.075
Sch.W. Oak	0.007	0.012	0.04	0.5	0.025	0.065



Sophie Zechmeister-Boltenstern
Katharina Keiblinger
BFW, Vienna

⇒ increase in protease-, cellulase- and xylanase-activities from Feb to May confirm semi-quantitative proteome data

PHOSPHOLIPID FATTY ACIDS (PLFA) ANALYSES

- ⇒ increase of the fungal community from Feb to May at Ort and Schottenwald (high in Mn and P) corresponds well to the semi-quantitative proteome analysis
- ⇒ no temporal changes in the community at Achenkirch and even a decrease in the fungal/bacterial ratio at Klausen-Leopoldsdorf (low in P and Mn); similar trends were reflected in our unique spectral counts



CONCLUSIONS – COMMUNITY STRUCTURE

- ⇒ generally, the **same taxonomic groups** were found **at all sampling sites & litter types**
- ⇒ the dominant taxa were **plants, fungi, and bacteria**
- ⇒ while **plant proteins decreased**, fungal and bacterial proteins increased from Feb to May at sampling sites with high P and Mn
- ⇒ a **lower number of microbial spectral counts** were observed **in samples with low P and Mn concentration** or of **structurally inert leafs** (oak); especially bacterial growth seems to be limited by phosphorus
- ⇒ while the composition of the **fungal community appeared unchanged** between Feb and May, the **proteobacterial community** seemed to **shift from δ - and γ - towards α - and β -proteobacteria**



CONCLUSIONS – COMMUNITY FUNCTION

- ⇒ under aerobic conditions **fungi seem to be the main degraders of beech litter**
- ⇒ **bacteria are present, but seem not to degrade the litter** (cheaters); they might play a more important role in anaerobic microniches or at later time points of the degradation process
- ⇒ **less litter degradation enzymes** were expressed **when either P or Mn were limited** (and thus fungal growth was limited)
- ⇒ **more fungal phosphatases** were expressed **at lower P concentrations**

The presence of a certain taxonomic group does not necessarily mean that it participates in a particular functional process, i.e. litter decomposition